A Genome-Scale Investigation of Incongruence in Culicidae Mosquitoes

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Abstract

Comparison of individual gene trees in several recent phylogenomic studies from diverse lineages has revealed a surprising amount of topological conflict or incongruence, but we still know relatively little about its distribution across the tree of life. To further our understanding of incongruence, the factors that contribute to it and how it can be ameliorated, we examined its distribution in a clade of 20 Culicidae mosquito species through the reconstruction and analysis of the phylogenetic histories of 2,007 groups of orthologous genes. Levels of incongruence were generally low, the three exceptions being the internodes concerned with the branching of Anopheles christyi, with the branching of the subgenus Anopheles as well as the already reported incongruence within the Anopheles gambiae species complex. Two of these incongruence events (A. gambiae species complex and A. christyi) are likely due to biological factors, whereas the third (subgenus Anopheles) is likely due to analytical factors. Similar to previous studies, the use of genes or internodes with high bootstrap support or internode certainty values, both of which were positively correlated with gene alignment length, substantially reduced the observed incongruence. However, the clade support values of the internodes concerned with the branching of the subgenus Anopheles as well as within the A. gambiae species complex remained very low. Based on these results, we infer that the prevalence of incongruence in Culicidae mosquitoes is generally low, that it likely stems from both analytical and biological factors, and that it can be ameliorated through the selection of genes with strong phylogenetic signal. More generally, selection of genes with strong phylogenetic signal may be a general empirical solution for reducing incongruence and increasing the robustness of inference in phylogenomic studies.

Key words: maximum likelihood, gene tree, bootstrap support (BS), bipartition, internode certainty (IC).

Recent advances in DNA sequencing technologies provide great opportunities for using genome-scale data to reconstruct phylogenetic history (Rokas and Abbott 2009; Hittinger et al. 2010; Faircloth et al. 2012; Lemmon et al. 2012). However, recent phylogenomic studies in diverse taxonomic groups, including plants (Zhong et al. 2013; Wickett et al. 2014), fungi (Hess and Goldman 2011; Salichos and Rokas 2013), and animals (Song et al. 2012; Jarvis et al. 2014), have shown that a large number of individual gene trees are topologically incongruent with each other. For example, a recent analysis of 1,070 orthologs from 23 yeast genomes identified 1,070 distinct gene trees, which were all incongruent with the phylogeny inferred from concatenation analysis (Salichos and Rokas 2013). Surprisingly, nearly half the internodes of the yeast phylogeny exhibited very low internode certainty (IC) values (Salichos and Rokas 2013), a measure of topological conflict (Salichos and Rokas 2013; Salichos et al. 2014). Similarly, the analysis of 32 plant taxa found 182 distinct topologies in a set of 184 gene trees (Zhong et al. 2013), and the analysis of 447 nuclear genes from 37 mammal species revealed 440 distinct topologies (Song et al. 2012).

Incongruence between gene trees can stem from analytical or biological factors. A wide variety of analytical factors can lead to failure to accurately infer a gene tree; these can be either due to stochastic error (e.g., insufficient sequence length or taxon samples) or due to systematic error (in case of departure from model assumptions, Jeffroy et al. 2006). In contrast, a number of biological factors can lead to gene trees that are actually distinct from each other and from the species tree. Examples of biological factors include incomplete lineage sorting (ILS), hidden paralogy, horizontal gene transfer, as well as gene duplication and loss, recombination and natural selection (Galtier and Daubin 2008; Degnan and Rosenberg 2009; Fontaine et al. 2015).
caused by analytical factors can be potentially reduced by some data filtering approaches, such as using genes with high phylogenetic information content (Dell’Ampio et al. 2014), slowly evolving genes (Betancur-R et al. 2014), genes with stationary base composition (Romiguier et al. 2013), genes with strong phylogenetic signals (Salichos and Rokas 2013), as well as internode-specific genes (Chen et al. 2015), incongruence stemming from biological factors cannot (Fontaine et al. 2015; Nater et al. 2015; Suh et al. 2015).

Although conflict between gene trees has been reported in analyses of phylogenomic data matrices from diverse plant, fungal, and animal taxa, we still know relatively little about the distribution of incongruence across the tree of life. Mosquitoes in the genus Anopheles represent an excellent lineage for investigating incongruence for two reasons. First, the draft genomes of 16 anophelines from Africa, Asia, Europe, and South America, representing a variety of geographic locations and ecological conditions and a range of evolutionary distances from each other, were recently sequenced (Neafsey et al. 2015). In addition to these 16 newly sequenced Anopheles genomes, the genomes of two additional Anopheles species, namely A. gambiae (Holt et al. 2002) and Anopheles darlingi (Marinotti et al. 2013), as well as the genomes of two other species belonging to the subfamily Culicinae, namely Aedes aegypti (Nene et al. 2007) and Culex quinquefasciatus (Arensburger et al. 2010), are also available.

The second reason is the demonstrated presence of incongruence in the Anopheles phylogeny, particularly within the Anopheles gambiae species complex (Besansky et al. 1994; Hittinger et al. 2010; Fontaine et al. 2015). Specifically, a genome-wide investigation of the relationships between the five species belonging to the A. gambiae complex, namely A. gambiae, Anopheles arabiensis, Anopheles quadriannulatus, Anopheles melas, and Anopheles merus, reported extensive introgression (Fontaine et al. 2015), prompting the authors of an associated commentary to ponder whether the notion of a bifurcating species phylogeny is a meaningful way to describe the evolutionary relationships among species in the complex (Clark and Messer 2015). Remarkably, it appears that the topology inferred from concatenation analysis, albeit strongly supported, is likely incorrect (Fontaine et al. 2015). This very high degree of incongruence raises the question on whether it is localized between species in the A. gambiae complex or whether it is also present in other parts of the Anopheles phylogeny.

Low Levels of Incongruence in Culicidae Phylogeny

In this study, we assembled a data set of 2,007 groups of orthologous genes (henceforth referred to simply as genes) from 20 Culicidae mosquito genomes (table 1). Maximum likelihood (ML) concatenation analysis of the 2,007-gene data matrix produced a species phylogeny in which all internodes exhibited 100% bootstrap support (BS) (fig. 1). Summarizing the 2,007 gene trees into an extended Majority Rule Consensus (eMRC) phylogeny or using them as input to construct a coalescent-based species phylogeny resulted in topologies that were identical to the concatenation phylogeny (fig. 1). Interestingly, 12 out of 17 internodes in the eMRC phylogeny had a gene-support frequency (GSF) of greater than 80%. Two of the remaining five internodes are associated with the branchings of Anopheles schristyi (GSF = 53%) and subgenus Anopheles (GSF = 62%), respectively, whereas the other three internodes show very low GSF values (33–43%) and all reside within the A. gambiae complex (fig. 1).

One thousand one hundred twenty-six of the 2,007 gene trees are unique, which means that about half of the gene trees do not agree (by at least one internode) with each other, or with species phylogeny supported by concatenation, eMRC, and coalescent-based approaches. The average normalized Robinson–Foulds (Robinson and Foulds 1981) tree distance between the 2,007 gene trees and the species phylogeny (0.21) was lower than that generated by an all-pairs comparison between the 2,007 gene trees (0.29) (fig. 2).

To quantify incongruence, we used IC which evaluates support for a given internode according its frequency in a given set of trees jointly with that the most prevalent conflicting bipartitions in the same set of trees (Salichos and Rokas 2013; Salichos et al. 2014). Examination of the eMRC phylogeny showed that 10 out of 17 internodes had IC values equal or greater than 0.70 and another two values greater than 0.55. The remaining five internodes, namely the branchings of A. christyi and subgenus Anopheles as well as the internodes within the A. gambiae species complex, had IC values less than 0.25. The branching of A. christyi depicts the bipartition (A. gambiae complex, A. christyi) (GSF = 53, IC = 0.08; fig. 1), which conflicts with the bipartition ([A. christyi, A. epiroticus], [remaining 18 species]), whose GSF is 27, yielding an IC value of 0.08. The branching of subgenus Anopheles depicts the bipartition (subgenus Cellia, subgenus Anopheles) (GSF = 62, IC = 0.19; fig. 1), which conflicts with the bipartition ([Anopheles albimanus, A. darlingi, Anopheles atroparvus, Anopheles sinensis], [remaining 16 species]), whose GSF is 21, yielding an IC value of 0.19.

The incongruence observed in internodes within the A. gambiae complex is much higher than the rest of the Culicidae phylogeny. The average GSF in the three internodes within the complex is 37.33, a value much smaller than the average GSF of 87.86 that is observed in the rest of the Culicidae phylogeny (fig. 1 and table 1). Similarly, the IC values of internodes in the A. gambiae complex rank first, third, and fifth lowest among the 17 internodes in the Culicidae phylogeny (fig. 1).
Table 1
The Effect of Using Genes and Bipartitions with Strong Phylogenetic Signal on the Culicidae Phylogeny

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Treatment Details</th>
<th>Average GSF</th>
<th>TC</th>
<th>RTC</th>
<th>Number of Internodes with Increased GSF</th>
<th>Number of Internodes with Decreased GSF</th>
<th>Number of Internodes with Increased IC</th>
<th>Number of Internodes with Decreased IC</th>
</tr>
</thead>
<tbody>
<tr>
<td>Default analysis</td>
<td>Default analysis</td>
<td>87.86</td>
<td>10.80</td>
<td>0.64</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td>Selection of genes whose ML trees have high average BS</td>
<td>Genes with average BS ≥ 70% (1,818 genes)</td>
<td>90.07</td>
<td>11.11</td>
<td>0.65</td>
<td>5</td>
<td>0</td>
<td>5</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>Genes with average BS ≥ 80% (1,379 genes)</td>
<td>92.14</td>
<td>11.70</td>
<td>0.69</td>
<td>11</td>
<td>0</td>
<td>10</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>Genes with average BS ≥ 90% (378 genes)</td>
<td>95.29</td>
<td>12.77</td>
<td>0.75</td>
<td>13</td>
<td>0</td>
<td>14</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>Genes with average BS ≥ 95% (66 genes)</td>
<td>96.43</td>
<td>13.02</td>
<td>0.77</td>
<td>15</td>
<td>0</td>
<td>14</td>
<td>1</td>
</tr>
<tr>
<td>Selection of genes whose ML trees have high TC</td>
<td>Using only the 1,818 genes with the highest TC</td>
<td>89.93</td>
<td>11.14</td>
<td>0.66</td>
<td>4</td>
<td>0</td>
<td>5</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>Using only the 1,379 genes with the highest TC</td>
<td>92.07</td>
<td>11.68</td>
<td>0.69</td>
<td>11</td>
<td>0</td>
<td>9</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>Using only the 378 genes with the highest TC</td>
<td>95.64</td>
<td>12.81</td>
<td>0.75</td>
<td>14</td>
<td>0</td>
<td>13</td>
<td>1</td>
</tr>
<tr>
<td></td>
<td>Using only the 66 genes with the highest TC</td>
<td>96.21</td>
<td>12.91</td>
<td>0.76</td>
<td>15</td>
<td>0</td>
<td>14</td>
<td>1</td>
</tr>
<tr>
<td>Selection of bipartitions with high BS in the ML trees of genes</td>
<td>Using only bipartitions that have ≥ 70% BS</td>
<td>NA</td>
<td>12.40</td>
<td>0.73</td>
<td>NA</td>
<td>NA</td>
<td>14</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>Using only bipartitions that have ≥ 80% BS</td>
<td>NA</td>
<td>12.88</td>
<td>0.76</td>
<td>NA</td>
<td>NA</td>
<td>14</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>Using only bipartitions that have ≥ 90% BS</td>
<td>NA</td>
<td>13.24</td>
<td>0.78</td>
<td>NA</td>
<td>NA</td>
<td>13</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>Using only bipartitions that have ≥ 95% BS</td>
<td>NA</td>
<td>13.34</td>
<td>0.78</td>
<td>NA</td>
<td>NA</td>
<td>13</td>
<td>0</td>
</tr>
</tbody>
</table>

Note.—The columns correspond to: the specific filtering of genes or bipartitions with strong phylogenetic signal tested (treatment and treatment details), the average GSF of the internodes of the Culicidae eMRC phylogeny (average GSF), the TC of the Culicidae eMRC phylogeny, the RTC of the Culicidae eMRC phylogeny, the numbers of internodes of the Culicidae eMRC phylogeny in which GSF increases or decreases by more than 3%, and the numbers of internodes of the Culicidae eMRC phylogeny in which IC increases or decreases by more than 0.03. As the maximum value of IC for a given internode is 1, the maximum value of TC for a given phylogeny is the number of internodes, which in this case is 17. In the analyses concerned with the use of bipartitions, only those bipartitions that displayed BS greater or equal to 70%, 80%, 90%, or 95% in the ML trees of the 2,007 genes were used to construct eMRC phylogenies, which were then compared with the default analysis. NA, not applicable.
To measure the degrees of conflict for every internode, IC can be more informative than GSF (Salichos and Rokas 2013; Salichos et al. 2014). For example, the placement of *Anopheles stephensi* and the placement of *Anopheles funestus* received 93% and 96% GSF, whereas their ICs were 0.75 and 0.93, respectively. This difference in the IC values of the two internodes despite similar GSF values is a result of the secondary conflicting signal difference. Specifically, whereas the most prevalent conflicting bipartition to the placement of *A. stephensi* has a GSF of 4%, the most prevalent conflict to the placement of *A. funestus* has a GSF of only 1%.

**Using Genes with Strong Phylogenetic Signal Reduces Incongruence**

To test whether using genes with stronger phylogenetic signal can reduce incongruence, we analyzed four data sets comprising genes whose ML trees had average BS values across all internodes greater than or equal to 70% (1,818 genes), 80% (1,379 genes), 90% (378 genes), or 95% (66 genes), and four data sets comprising the 1,818, 1,379, 378, or 66 genes whose ML trees had the highest tree certainty (TC) values. Note that gene selection was solely based on the strength of phylogenetic signal exhibited in their gene trees (measured by BS or TC) without any consideration to the topology supported. The concatenation analysis as well as the eMRC analysis was redone each time when the new data set was selected. We found that the GSF and IC values of the vast majority of internodes increased as the stringency of the BS and TC filters increased (supplementary table S2, Supplementary Material online), suggesting that selecting genes with high average BS or high TC significantly reduced incongruence in the Culicidae phylogeny (table 1).

We also tested whether using internodes with high BS can reduce the incongruence by extracting bipartitions with BS values greater than or equal to 70%, 80%, 90% or 95% from every ML tree of the 2,007 genes and then used them to construct the eMRC phylogenies. Importantly, the use of highly supported bipartitions allows one to quantify a given internode’s IC from only the subset of bipartitions that highly support or conflict with that internode (Salichos and Rokas 2013; Salichos et al. 2014). Compared to the phylogeny of figure 1, this practice significantly increased IC values for ≥ 13 internodes (table 1 and supplementary table S2, Supplementary Material online).

Even though the GSF and IC values of the internodes concerned with the branching of the subgenus *Anopheles*, with...
the branchings within the A. gambiae species complex, and
with the branching of A. christyi, increased as the stringency
of the BS and TC filters increased, the values themselves were still
lower even for the strictest filters (supplementary table S2,
Supplementary Material online). Thus, incongruence in these
internodes may be the result of biological factors such as ILS,
gene duplication and loss, or introgression (Fontaine et al.
2015).

The Relationship between
Incongruence and Gene
Alignment Length

Both average BS as well as TC values are positively correlated
with genes’ alignment lengths (alignment length vs. BS, \( r = 0.50, P \) value < 2.2e-16; alignment length vs. TC, \( r = 0.56, P \) value < 2.2e-16; supplementary fig. S1, Supplementary Material online). How does incongruence behave if we use genes with the same lengths? To resolve this question, we created a new data matrix that contained only the first 999bp of the sequence alignment of the 1,340 genes that were 999bp or longer (genes with shorter alignment lengths were excluded) and re-analyzed levels of incongruence in the Culicidae phylogeny. The results are quite similar to the results from the 2,007-gene data matrix; 12 of the 17 internodes exhibit high GSF and IC values, whereas internodes within the A. gambiae species complex as well as internodes associated with the placement of A. christyi and the subgenus Anopheles show low GSF and very low IC values (fig. 3).

Thus, although the average BS and TC values were positively correlated with gene alignment length, using loci that have the same alignment lengths does not appear to substantially decrease or increase the incongruence present in this phylogenomic data matrix.

We also examined whether the selection of genes or bipartitions with strong phylogenetic signal in this set of 1,340 alignment length-standardized genes reduced incongruence. We tested three data sets comprising genes whose ML trees showed average BS values across all internodes that were greater than or equal to 70% (1,138 genes), 80% (603 genes) or 90% (45 genes) (no gene had average BS greater or equal to 95%), and three data sets comprising the 1,138, 603 or 45 genes whose trees had the highest TC. Almost all the GSF and IC values of every internode increased as the value of the BS or TC filter increased (table 2 and supplementary table S3, Supplementary Material online). Using genes or internodes with high BS or IC values also significantly reduced the observed incongruence (table 2 and supplementary table S3, Supplementary Material online). Similarly, selecting internodes with high BS decreased incongruence by extracting only those bipartitions that display BS values greater than or equal to 70%, 80%, 90% or 95% from every one of the 1,340 genes’ ML trees. This practice significantly increased IC values for \( \geq 14 \) internodes relative to the phylogeny of figure 3 (table 2 and supplementary table S3, Supplementary Material online). However, the GSF and IC values of the internodes concerned with the branching of the subgenus Anopheles, the branching of A. christyi, as

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**Fig. 2.**—The distribution of the agreement between the bipartitions present in the 2,007 individual gene trees and the concatenation phylogeny, measured using the normalized Robinson–Foulds tree distance. The phylogeny of the 20 Culicidae species analyzed in this study is unrooted and contains 17 nontrivial bipartitions.
Conclusion

In summary, analyses of a 2,007-gene phylogenomic data matrix from 20 Culicidae mosquito genomes showed that incongruence was low and localized to specific branches. Similar to previous studies, the use of genes or bipartitions with strong phylogenetic signal (quantified either through the use of BS or IC values) substantially reduced the observed incongruence. However, the GSF and IC values of the internodes concerned with the branchings of *A. christyi*, the subgenus *Anopheles*, as well as with branchings within the *A. gambiae* species complex remained very low. Combined with the observation that many of the *A. gambiae* species complex internodes are very short, the observed incongruence is consistent with previous inferences of extensive introgression within the *A. gambiae* species complex (Fontaine et al. 2015). Short internode length makes biological factors, such as ILS or introgression, the most likely explanation for the incongruence observed in the branching of *A. christyi*. In contrast, the internode associated with the branching of the subgenus *Anopheles* and the subgenus *Celia* is much longer suggesting that this incongruence is more likely to be due to analytical factors. Very similar results were obtained with a 1,340-gene phylogenomic data matrix in which all genes had the same length, arguing that the well-known correlation between alignment length and phylogenetic signal did not have a major influence on phylogenetic reconstruction in this lineage. What’s more, they add to the body of evidence (Salichos and Rokas 2013) showing that the selection of genes with strong phylogenetic signal can reduce incongruence and increase the robustness of phylogenetic inference. Thus, this strategy may be a general empirical solution for ameliorating incongruence in phylogenomic studies.

Materials and Methods

Data Matrix Construction

We used the complete sets of annotated orthology data of 20 Culicidae mosquito genomes (supplementary table S1, Supplementary Material online) from http://cegg.unige.ch/orthodbmoz2 (Neafsey et al. 2015). We selected 2,008 single-copy genes which that contained sequences from all 20 species as our initial data set. The nucleotide sequences...
Table 2
The Effect of Using Genes and Bipartitions with Strong Phylogenetic Signal on the Culicidae Phylogeny Based on the First 999 bp of Every Gene’s Alignment

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Treatment Details</th>
<th>Average GSF</th>
<th>TC</th>
<th>RTC</th>
<th>Number of Internodes with Increased GSF</th>
<th>Number of Internodes with Decreased GSF</th>
<th>Number of Internodes with Increased IC</th>
<th>Number of Internodes with Decreased IC</th>
</tr>
</thead>
<tbody>
<tr>
<td>1,340 genes</td>
<td>First 999 bp sequence of every gene (1,340 genes)</td>
<td>84.43</td>
<td>9.98</td>
<td>0.59</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td>Selection of genes whose genes with average BS ≥ 70% (1,138 genes)</td>
<td>87.36</td>
<td>10.34</td>
<td>0.61</td>
<td>8</td>
<td>0</td>
<td>5</td>
<td>0</td>
<td></td>
</tr>
<tr>
<td>ML trees have high average BS</td>
<td>91.00</td>
<td>11.17</td>
<td>0.66</td>
<td>14</td>
<td>0</td>
<td>10</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td>Selection of genes whose ML trees have high TC</td>
<td>87.29</td>
<td>10.34</td>
<td>0.60</td>
<td>10</td>
<td>0</td>
<td>7</td>
<td>0</td>
<td></td>
</tr>
<tr>
<td>Selection of bipartitions with high BS in the ML trees of genes</td>
<td>90.47</td>
<td>11.16</td>
<td>0.63</td>
<td>13</td>
<td>0</td>
<td>10</td>
<td>0</td>
<td></td>
</tr>
<tr>
<td>Selection of genes whose ML trees have high TC</td>
<td>94.86</td>
<td>12.23</td>
<td>0.72</td>
<td>14</td>
<td>0</td>
<td>13</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td>Selection of genes whose ML trees have high TC</td>
<td>94.86</td>
<td>12.23</td>
<td>0.72</td>
<td>14</td>
<td>0</td>
<td>13</td>
<td>1</td>
<td></td>
</tr>
</tbody>
</table>

Note.—The columns correspond to the specific filtering of genes or bipartitions with strong phylogenetic signal tested (treatment and treatment details), the average GSF of the internodes of the Culicidae eMRC phylogeny (average GSF), the TC of the Culicidae eMRC phylogeny, the RTC of the Culicidae eMRC phylogeny, the numbers of internodes of the Culicidae eMRC phylogeny in which GSF increases or decreases by more than 3%, and the numbers of internodes of the Culicidae eMRC phylogeny in which IC increases or decreases by more than 0.03. As the maximum value of IC for a given internode is 1, the maximum value of TC for a given phylogeny is the number of internodes, which in this case is 17. In the analyses concerned with the use of bipartitions, only those bipartitions that displayed BS greater or equal to 70%, 80%, 90%, or 95% in the ML trees of the 1,340 genes were used to construct eMRC phylogenies, which were then compared with the default analysis. NA, not applicable.

Gene Alignment

We aligned all genes using the MAFFT software, version 7.182 (KatOH and TOH 2008) based on their amino acid sequence. For the codon sequence alignment of each gene, the un-rooted eMRC phylogeny that consisted of those genes with a 999 bp long alignment and does not contain any missing data was translated to amino acids. A series of different data sets were constructed using custom Perl scripts. To test how incongruence varied independent of gene alignment data size, we used PAL2NAL (Suyama et al. 2008) to translate amino acid alignments intended to be corresponding to data size, then, we used RAxML (Stamatakis 2014), under the GTRGAMMA model and with the values of the nucleotide base frequencies fixed to “observed” and those of the substitution rate parameters estimated from the data (raxmlHPC-PTHREADS-SSE3 -T 8 -f a -x 12345 -p 12345 -N 100 -m GTRGAMMA -s ALIGNMENT -n NAME).
version 3.696 (PHYLIP; J. Felsenstein, University of Washington, Seattle; http://evolution.genetics.washington.edu/phylip.html). The eMRC phylogeny of bipartitions with high BS was constructed using custom Perl scripts. As the divergence of Culicinae and Anophelinae lineages is well established, all phylogenies shown in figures have been midpoint rooted at the internode that separates these two lineages for easier visualization.

The coalescent species phylogeny was estimated using 100 replicates of multi-locus bootstrapping in ASTRAL (Mirarab and Warnow 2015) (java -Xmx36000M -jar astral.4.7.jar -i TREECOLLECTION -o OUTPUT -b BS_PATH -r 100), and using the online version of the STAR software with 100 rooted bootstrap replicates of every gene (Liu et al. 2009; http://bioinformatics.publichealth.uga.edu/SpeciesTreeAnalysis/STAR/STAR.php).

**Data Availability**

All data and analyses described in this study are deposited at Figshare under the accession 10.6084/m9.figshare.1566851.

**Supplementary Material**

Supplementary figure S1 and tables S1–S3 are available at Genome Biology and Evolution online (http://www.gbe.oxfordjournals.org).

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**Literature Cited**


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