

SUMMER INTERNSHIP PROJECT REPORT

submitted by

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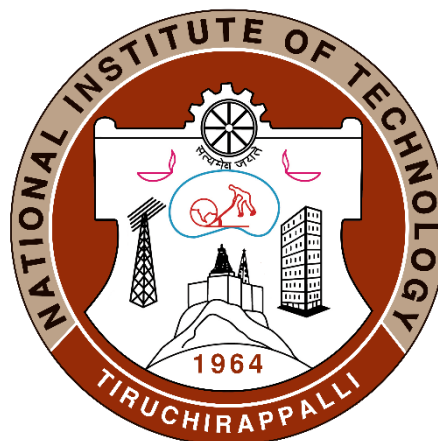
Roll no: **102118064**

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Organisation Profile

The Department of Biotechnology at IIT Madras was founded in 2004 with a vision to be recognised as a department of international reputation with a strong interdisciplinary research and teaching base in biological sciences and engineering and active collaboration with industries and healthcare institutions. The department is housed in the 'Bhupat and Jyoti Mehta School of Biosciences'.

Modern biotechnology finds applications in a variety of domains, such as in the manufacture of high-technology pharmaceuticals and the making of enzymes for the detergents we use for washing our clothes. With expanding knowledge of the cell and its functioning at the molecular level, biotechnologists are now seeking answers to prevent life-threatening diseases, meet the energy and food requirements of an ever-growing population, and solve the environmental problems that threaten our very existence on this planet. The diversity of challenges that biotechnology tackle is reflected in the research activities of the faculty of Biotechnology at IIT Madras. Although there are many different research groups in the department, their research generally falls into one or more of the following categories: medical biotechnology related to cancer and cardiovascular aspects, chemical biology, computational biology and bioprocesses.

It carries out collaborative research with other departments within the institute and other institutes in India and abroad. In the past few years, their research contributions through journal publications have been significant, and they have also filed for several patents.

Background of Study

It is widely known that the population of the world multiplies exponentially with every passing generation. This results in the increased requirement of technology and power, which demands massive quantities of fuel. The primary fuels used are products of petroleum, which are non-renewable sources of energy. They also produce carbon dioxide after combustion, which results in what is known as the greenhouse effect. This has started to burn a hole in the ozone layer in the stratosphere and cause alarming climate change.

This calls for the need for renewable sources of energy, which also need to have little to no negative impact on the atmosphere. There are various options like solar power, wind power, hydroelectric power and tidal power. But it is important to study further and find more sources of such power in unlikely places and learn to harness and exploit these resources.

Acinetobacter is a group of bacteria commonly found in the environment, like in soil and water. It is a genus of gram-negative bacteria that are oxidase-negative, exhibit twitching motility and occurs in pairs under magnification. They are important soil organisms that contribute to the mineralisation of aromatic compounds.

Most strains of *Acinetobacter* grow well on MacConkey agar without salt. Bacteria of the genus *Acinetobacter* are known to form intracellular inclusions of polyhydroxyalkanoates under certain environmental conditions (e.g. lack of elements such as phosphorus, nitrogen, or oxygen combined with an excessive supply of carbon sources).

Different species of bacteria in this genus can be identified using fluorescence-lactose-denitrification to find the amount of acid produced by the metabolism of glucose. The other reliable identification test at the genus level is the chromosomal DNA transformation assay. In this assay, a naturally competent tryptophan auxotrophic mutant of *Acinetobacter baylyi* (BD4 trpE27) is transformed with the total DNA of a putative *Acinetobacter* isolate, and the transformation mixture is plated on a brain heart infusion agar. The growth is then harvested after incubation for 24 h at 30 °C, plating on an *Acinetobacter* minimal agar (AMA), and incubating at 30 °C for 108 h. Growth on the AMA indicates a positive transformation assay.

This *A. baylyi* is modified through a process called adaptive laboratory evolution by growing on a mixture of monomers derived from lignin. Adaptive laboratory evolution (ALE) is a frequent method in biological studies to gain insights into the basic mechanisms of molecular evolution and adaptive changes that accumulate in microbial populations during long term selection under specified growth conditions. It is used to break down lignin to produce an eco-friendly source of energy. This study involves looking for the mutations that may have a role to play in the evolved strain and uptake of the lignin monomers. This can be validated by performing experiments in the laboratory.

Whole-genome sequencing (WGS) is a comprehensive method for analyzing entire genomes. Genomic information has been instrumental in identifying inherited disorders, characterizing the mutations that drive cancer progression, and tracking disease outbreaks. This method is used to analyse mutations and conclude the effects of said mutations. In this study, whole

genome sequence analysis has been done to identify the mutations undergone by the strain through the process of ALE. The various steps performed in this analysis can be broadly listed as follows:

- ◆ Genome assembly
- ◆ Ordering and viewing of the assembled contigs
- ◆ Genome annotation

These mutations are then studied to gauge the effects they produce in the strain. These effects can be validated by testing the strains in the laboratory.

Work Done

Whole-genome sequence analysis is the process of determining the entirety, or nearly the entirety, of the DNA sequence of the genome of an organism at a single time. This entails sequencing all of the organism's chromosomal and cellular DNA.

Bacterial whole-genome sequencing (WGS) is becoming a widely-used technique in research, clinical diagnostic, and public health laboratories. It enables high-resolution characterisation of bacterial pathogens in terms of properties that include antibiotic resistance, molecular epidemiology, and virulence.

This computational analysis involved the whole genome sequence analysis of *Acinetobacter baylyi*, and it was performed on <https://usegalaxy.org/>. The process was done in a few steps, which have been put together below.

1. Reference Genome

Downloaded the reference genome – *Acinetobacter* sp. ADP1 (GenBank: CR 543861.1)

2. FASTQ Summary Statistics

Found the quality statistics of the wild type and evolved strain using FASTQ Summary Statistics tool [\[1\]](#) and obtained a tabular file containing the following as output:

- ◆ column = column number (1 to 36 for a 36-cycles read Solexa file)
- ◆ count = number of bases found in this column.
- ◆ min = Lowest quality score value found in this column.
- ◆ max = Highest quality score value found in this column.
- ◆ sum = Sum of quality score values for this column.
- ◆ mean = Mean quality score value for this column.
- ◆ Q1 = 1st quartile quality score.
- ◆ med = Median quality score.
- ◆ Q3 = 3rd quartile quality score.
- ◆ IQR = Inter-Quartile range (Q3-Q1).
- ◆ LW = 'Left-Whisker' value (for boxplotting).
- ◆ rW = 'Right-Whisker' value (for boxplotting).
- ◆ outliers = Scores falling beyond the left and right whiskers (comma separated list).
- ◆ A_Count = Count of 'A' nucleotides found in this column.
- ◆ C_Count = Count of 'C' nucleotides found in this column.
- ◆ G_Count = Count of 'G' nucleotides found in this column.
- ◆ T_Count = Count of 'T' nucleotides found in this column.
- ◆ N_Count = Count of 'N' nucleotides found in this column.
- ◆ Other_Nucs = Comma separated list of other nucleotides found in this column.
- ◆ Other_Count = Comma separated count of other nucleotides found in this column.

3. *Quality assessment of raw reads*

Performed quality assessment of raw reads using FASTQC tool [\[2\]](#) and obtained a basic text and a HTML file containing the following results:

- ◆ Basic Statistics
- ◆ Per base sequence quality
- ◆ Per sequence quality scores
- ◆ Per base sequence content
- ◆ Per base GC content
- ◆ Per sequence GC content
- ◆ Per base N content
- ◆ Sequence Length Distribution
- ◆ Sequence Duplication Levels
- ◆ Overrepresented sequences
- ◆ Kmer Content

All except the basic statistics and overrepresented sequences were plots.

4. *Trimming of raw reads*

Trimmed the raw reads using Trimmomatic flexible read trimming tool [\[3\]](#) for Illumina NGS data, based on the quality assessment to remove reads of low quality and adapter sequences and performed the following steps to obtain trimmed files:

- ◆ ILLUMINACLIP Step with TruSeq2 adapter sequences (Single-ended, for Illumina GAII)
- ◆ LEADING operation to keep bases of a minimum quality of 3
- ◆ TRAILING operation to keep bases of a minimum quality of 3
- ◆ SLIDINGWINDOW operation with average quality being 15 across 4 bases
- ◆ MINLEN operation to keep reads of minimum length 20

The following outputs were obtained:

- ◆ R1 and R2 paired reads of the wild type
- ◆ R1 and R2 unpaired reads of the wild type
- ◆ R1 and R2 paired reads of the evolved strain
- ◆ R1 and R2 unpaired reads of the evolved strain

5. *Mapping of genome to reference*

Mapped the sequence to reference genome after removing redundant data using Map with BWA tool. [\[4\]](#) [\[5\]](#) BWA is a software package for mapping low-divergent sequences against a large reference genome, such as the human genome. The bwa-aln algorithm is

designed for Illumina sequence reads up to 100bp. This tool wraps bwa-aln, bwa-samse and bwa-sampe modules of bwa read mapping tool:

- ◆ bwa aln - actual mapper placing reads onto the reference sequence
- ◆ bwa samse - post-processor converting suffix array co-ordinates into genome co-ordinates in SAM format for single reads
- ◆ bam sampe - post-processor for paired reads

The implementation takes fastq or BAM (unaligned BAM) datasets as input and produces output in BAM format, which can be further processed using various BAM utilities (BAMTools, SAMTools, Picard).

6. *Sorting the BAM dataset*

Sorted the BAM dataset using the SortSAM tool [\[6\]](#), which uses Picard documentation. Both input and output were in BAM format.

7. *Marking and removing duplicate reads*

Examined the aligned records in the BAM dataset given as input to locate duplicate molecules using the MarkDuplicates tool [\[6\]](#), which uses Picard documentation. All records are then written to the output file with duplicate records flagged. The outputs included the co-ordinate sorted BAM files and the metrics in a text file.

8. *Variant Calling*

Used FreeBayes, [\[7\]](#) a Bayesian genetic variant detector designed to find small polymorphisms, specifically single nucleotide polymorphisms (SNPs), insertions and deletions (indels), multiple nucleotide polymorphisms (MNPs) and complex events (composition insertion and substitution events) smaller than the length of a short-read sequencing alignment.

Provided some BAM dataset(s) and a reference sequence, FreeBayes will produce a VCF dataset describing SNPs, indels, and complex variants in samples in the input alignments.

By default, FreeBayes will consider variants supported by at least two observations in a single sample (-C) and also by at least 20% of the reads from a single sample (-F). These settings are suitable for low to high depth sequencing in haploid and diploid samples, but users working with polyploid or pooled samples may wish to adjust them depending on the characteristics of their sequencing data.

FreeBayes is capable of calling variant haplotypes shorter than a read length where multiple polymorphisms segregate on the same read. The maximum distance between polymorphisms phased in this way is determined by the --max-complex-gap, which defaults to 3bp. In practice, this can comfortably be set to half the read length.

Ploidy may be set to any level (-p), but by default, all samples are assumed to be diploid. FreeBayes can model per-sample and per-region variation in copy-number (-A) using a copy-number variation map.

FreeBayes can act as a frequency-based pooled caller and describe variants and haplotypes in terms of observation frequency rather than called genotypes. Allele observation counts will be described by AO and RO fields in the VCF output.

9. Building a database

Used the SnpEff build – genbank [\[8\]](#) command in the SnpEff build tool to create an SnpEff database. Output was in snpeffdb format.

10. Annotating variants

Annotated and predicted the effects of genetic variants using the SnpEff eff tool. [\[8\]](#)

A typical SnpEff use case would be:

- ◆ Input: The inputs are predicted variants (SNPs, insertions, deletions and MNPs). The input file is usually obtained as a result of a sequencing experiment, and it is usually in variant call format (VCF).
- ◆ Output: SnpEff analyses the input variants. It annotates the variants and calculates the effects they produce on known genes (e.g. amino acid changes). A list of effects and annotations that SnpEff can calculate can be found [here](#).

In a typical sequencing experiment, there will be many places in the genome where your sample differs from the reference genome. These are called "genomic variants" or just "variants".

11. Compiling mutations

Manually checked the positions of the reference genome to find out the type of gene, the product it coded for, and the change that occurred using a genome browser tool. [\[9\]](#) The data was compiled in a spreadsheet.

Outputs

1. FASTQ Summary Statistics

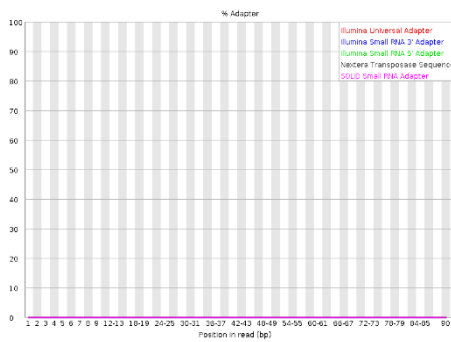
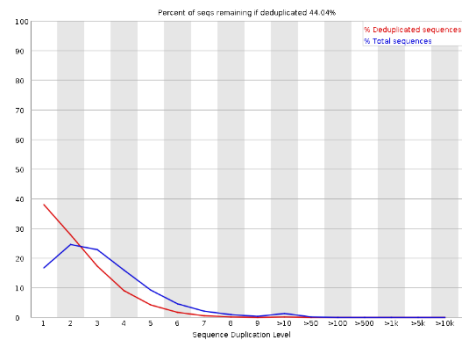
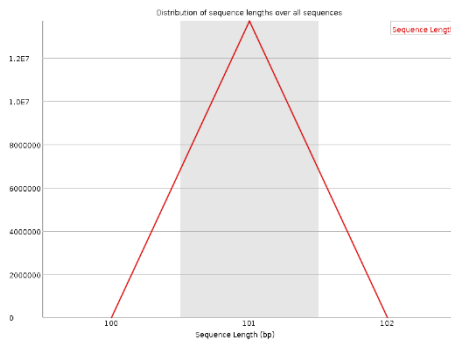
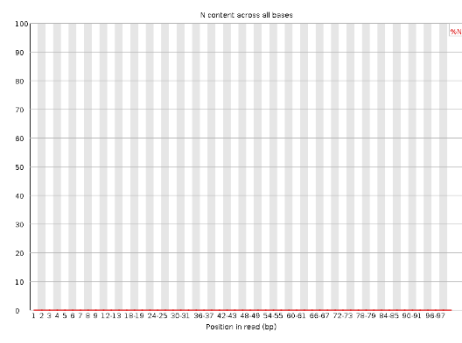
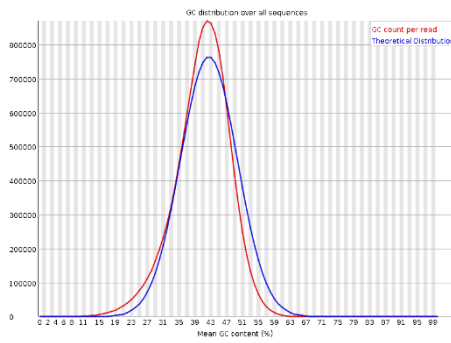
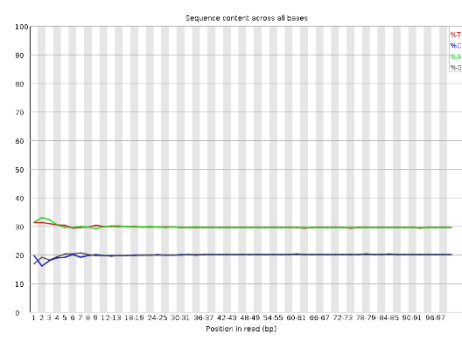
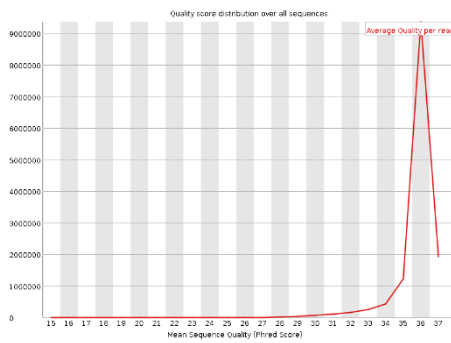
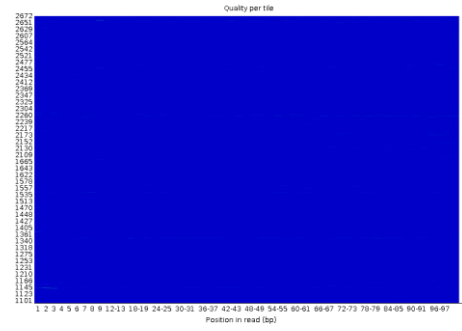
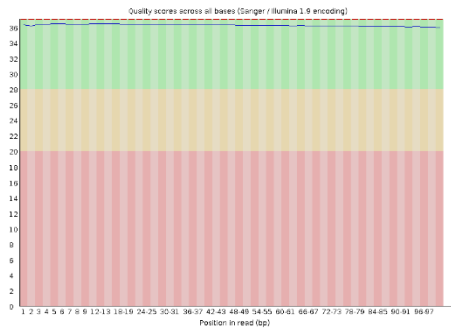
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67	13712696	11	37	4.97E+08	36.21256	37	37	37	0	37	37	11,25	4090248	2780596	2773835	4068017	0		
68	13712696	11	37	4.96E+08	36.16804	37	37	37	0	37	37	11,25	4080631	2774921	2792785	4064359	0		
69	13712696	11	37	4.96E+08	36.2033	37	37	37	0	37	37	11,25	4062779	2804112					

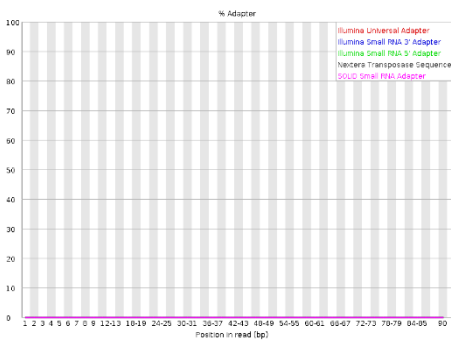
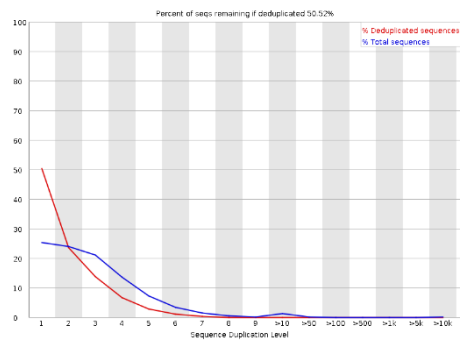
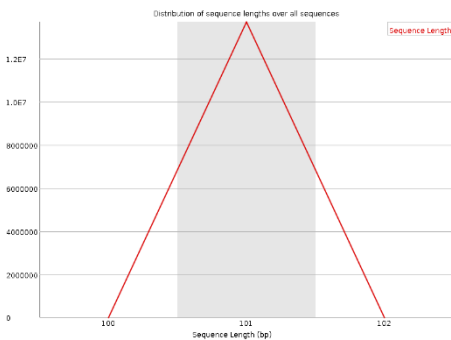
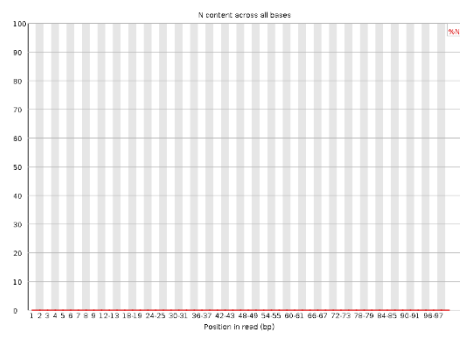
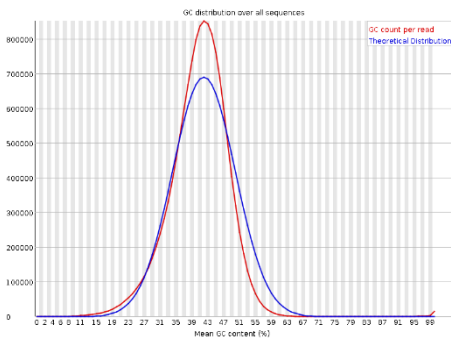
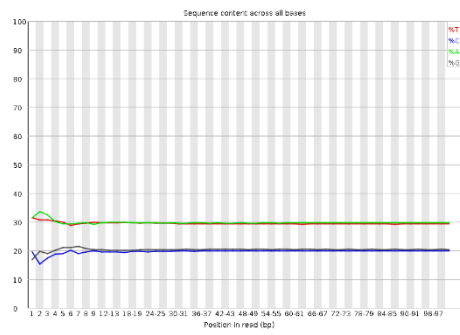
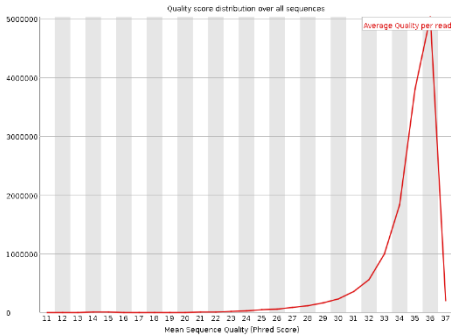
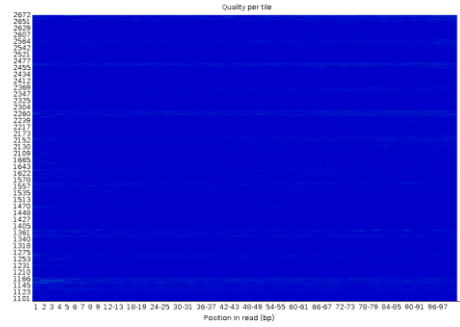
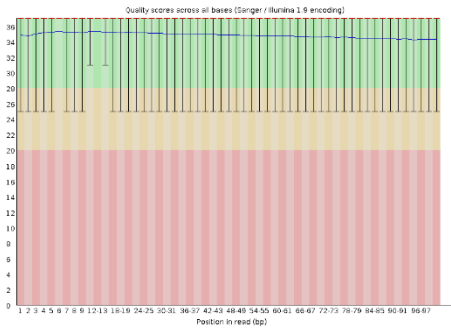
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2	13712696	2	37	4.77E+08	34.76612	37	37	37	0	37	37	2,11,25	4620088	2133611	2722317	4230688	5992		
3	13712696	2	37	4.81E+08	35.06586	37	37	37	0	37	37	2,11,25	4463674	2411276	2615241	4222491	14		
4	13712696	11	37	4.83E+08	35.19121	37	37	37	0	37	37	11,25	4164917	2598864	2711336	4117179	0		
5	13712696	11	37	4.84E+08	35.28636	37	37	37	0	37	37	11,25	4044945	2619915	2908593	4139243	0		
6	13712696	11	37	4.84E+08	35.30578	37	37	37	0	37	37	11,25	4050251	2784136	2904346	3973963	0		
7	13712696	11	37	4.83E+08	35.22262	37	37	37	0	37	37	11,25	4080477	2763169	2955168	4040882	0		
8	13712696	11	37	4.83E+08	35.22615	37	37	37	0	37	37	11,25	4099027	2691909	2856998	4064762	0		
9	13712696	11	37	4.83E+08	35.21868	37	37	37	0	37	37	11,25	4017050	2743239	2818114	4134293	0		
10	13712696	11	37	4.84E+08	35.28431	37	37	37	0	37	37	11,25	4103014	2707783	2813677	4088222	0		
11	13712696	11	37	4.84E+08	35.32254	37	37	37	0	37	37	11,25	4106600	2688211	2809774	4108111	0		
12	13712696	11	37	4.84E+08	35.32991	37	37	37	0	37	37	11,25	4108241	2718004	2793813	4092638	0		
13	13712696	11	37	4.84E+08	35.30304	37	37	37	0	37	37	11,25	4150627	2678962	2761644	4121463	0		
14	13712696	11	37	4.84E+08	35.28495	37	37	37	0	37	37	11,25	4150759	2663598	2788054	4110285	0		
15	13712696	11	37	4.83E+08	35.23449	37	37	37	0	37	37	11,25	4116318	2720075	2774760	4101543	0		
16	13712696	11	37	4.84E+08	35.26476	37	37	37	0	37	37	11,25	4131217	2689749	2765035	4126095	0		
17	13712696	11	37	4.83E+08	35.25716	37	37	37	0	37	37	11,25	4118199	2686055	2794073	4114369	0		
18	13712696	11	37	4.83E+08	35.25806	37	37	37	0	37	37	11,25	4085553	2737438	2788175	4101530	0		
19	13712696	11	37	4.82E+08	35.14295	37	37	37	0	37	37	11,25	4117932	2704792	2778690	4111282	0		
20	13712696	2	37	4.83E+08	35.22849	37	37	37	0	37	37	2,11,25	4111674	2694348	2812124	4094503	31		
21	13712696	11	37	4.83E+08	35.24118	37	37	37	0	37	37	11,25	4083539	2741641	2810532	4076984	0		
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23	13712696	11	37	4.83E+08	35.20218	37	37	37	0	37	37	11,25	4109961	2702127	2813668	4086940	0		
24	13712696	11	37	4.82E+08	35.14205	37	37	37	0	37	37	11,25	4086229	2745548	2815403	4065516	0		
25	13712696	11	37	4.82E+08	35.18521	37	37	37	0	37	37	11,25	4109814	2725050	2795828	4082004	0		
26	13712696	11	37	4.82E+08	35.11746	37	37	37	0	37	37	11,25	4098994	2713967	2822555	4077180	0		
27	13712696	2	37	4.82E+08	35.11418	37	37	37	0	37	37	11,25	4075717	2755334	2820601	4061044	0		
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29	13712696	11	37	4.81E+08	35.07686	37	37	37	0	37	37	11,25	4098215	2718646	2830450	4065385	0		
30	13712696	11	37	4.81E+08	35.09979	37	37	37	0	37	37	11,25	4071571	2762398	2827035	4051692	0		
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33	13712696	11	37	4.8E+08	34.98056	37	37	37	0	37	37	11,25	4072759	2768368	2832162	4039407	0		
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37	13712696	11	37	4.81E+08	35.04127	37	37	37	0	37	37	11,25	4103527	2746246	2807880	4055043	0		
38	13712696	2	37	4.8E+08	34.97932	37	37	37	0	37	37	2,11,25	4094446	2735813	2835128	4046987	322		
39	13712696	11	37	4.79E+08	34.96345	37	37	37	0	37	37	11,25	4073735	2773807	2831635	4033519	0		
40	13712696	11	37	4.8E+08	35.02246	37	37	37	0	37	37	11,25	4102993	2749038	2807249	4053416	0		
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42	13712696	11	37	4.79E+08	34.95226	37	37	37	0	37	37	11,25	4069424	2774614	2839150	4029508	0		
43	13712696	11	37	4.8E+08	35.00188	37	37	37	0	37	37	11,25	4098820	2753747	2810967	4049162	0		
44	13712696	11	37	4.79E+08	34.91069	37	37	37	0	37	37	11,25	4093836	2734406	2835341	4046563	0		
45	13712696	11	37	4.79E+08	34.92247	37	37	37	0	37	37	11,25	4073343	2772032	2834785	4032536	0		
46	13712696	11	37	4.79E+08	34.94176	37	37	37	0	37	37	11,25	4096698	2752578	2808567	4054853	0		
47	13712696	11	37	4.79E+08	34.90024	37	37	37	0	37	37	11,25	4092763	2739724	2839497	4040712	0		
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51	13712696	2	37	4.78E+08	34.85432	37	37	37	0	37	37	2,11,25	4074688	2778101	2832272	4027614	21		
52	13712696	11	37	4.78E+08	34.82732	37	37	37	0	37	37	11,25	4102025	2755965	2808327	4046379	0		
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61	13712696	11	37	4.76E+08	34.73416	37	37	37	0	37	37	11,25	4105094	2754263	2804668	4048671	0		
62	13712696	11	37	4.77E+08	34.76756	37	37	37	0	37	37	11,25	4098430	2741071	2833032	4040163	0		
63	13712696	11	37	4.76E+08	34.69583	37	37	37	0	37	37	11,25	4077548	2777771	2828746	4028631	0		
64	13712696	11	37	4.75E+08	34.66546	37	37	37	0	37	37	11,25	4100851	2755937	2810532	4045376	0		
65	13712696	11	37	4.76E+08	34.68046	37	37	37	0	37	37	11,25	4099147	2738773	2833633	4041143	0		
66	13712696	11	37	4.76E+08	34.69049	37	37	37	0	37	37	11,25	4076879	2775598	2829289	4030930	0		
67	13712696	11	37	4.76E+08	34.68559	37	37	37	0	37	37	11,25	4104939	2751990	2806714	4049053	0		
68	13712696	11	37	4.75E+08	34.64206	37	37	37	0	37	37	11,25	4102315	2735283	2831913	4043185	0		
69	13712696	11	37	4.76E+08	34.65422	37	37</												

row	column	count	min	max	sum	mean	Q1	med	Q3	IQR	IW	rW	outliers	A_Count	C_Count	G_Count	T_Count	N_Count	other_bases	other_base_count
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3	15251046	2	37	5.52E+08	36.20007	37	37	37	0	37	37	2,11,25	4820442	2775250	2797657	4857669	28			
4	15251046	11	37	5.54E+08	36.31029	37	37	37	0	37	37	2,11,25	4593562	2909478	3017520	4730486	0			
5	15251046	2	37	5.55E+08	36.40696	37	37	37	0	37	37	2,11,25	4538951	2952859	3160254	4598880	2			
6	15251046	11	37	5.56E+08	36.43477	37	37	37	0	37	37	2,11,25	4520503	3125605	3152568	4452370	0			
7	15251046	11	37	5.56E+08	36.43455	37	37	37	0	37	37	2,11,25	4537368	3014138	3177217	4522323	0			
8	15251046	11	37	5.56E+08	36.44148	37	37	37	0	37	37	2,11,25	4523366	3093799	3079755	4554126	0			
9	15251046	11	37	5.56E+08	36.44065	37	37	37	0	37	37	2,11,25	4453265	3114696	3064323	4618762	0			
10	15251046	11	37	5.56E+08	36.44669	37	37	37	0	37	37	2,11,25	4558511	3061572	3064143	4566820	0			
11	15251046	11	37	5.56E+08	36.46819	37	37	37	0	37	37	2,11,25	4560600	3036080	3055115	4599251	0			
12	15251046	11	37	5.56E+08	36.47541	37	37	37	0	37	37	2,11,25	4548828	3068085	3034191	4599942	0			
13	15251046	11	37	5.56E+08	36.46132	37	37	37	0	37	37	2,11,25	4587576	3032081	3018689	4612700	0			
14	15251046	11	37	5.56E+08	36.45678	37	37	37	0	37	37	2,11,25	4575374	3030259	3031424	4613989	0			
15	15251046	11	37	5.56E+08	36.44848	37	37	37	0	37	37	2,11,25	4546407	3074578	3024128	4605933	0			
16	15251046	11	37	5.56E+08	36.44907	37	37	37	0	37	37	2,11,25	4568912	3040122	3030406	4611606	0			
17	15251046	2	37	5.56E+08	36.46089	37	37	37	0	37	37	2,11,25	4560637	3039311	3045306	4605780	12			
18	15251046	11	37	5.56E+08	36.45963	37	37	37	0	37	37	2,11,25	4538308	3080053	3037851	4594834	0			
19	15251046	11	37	5.56E+08	36.45311	37	37	37	0	37	37	2,11,25	4568709	3045972	3042962	4593403	0			
20	15251046	11	37	5.56E+08	36.45394	37	37	37	0	37	37	2,11,25	4562187	3035419	3064656	4588784	0			
21	15251046	11	37	5.56E+08	36.44933	37	37	37	0	37	37	2,11,25	4540025	3076327	3059369	4573525	0			
22	15251046	11	37	5.56E+08	36.43303	37	37	37	0	37	37	2,11,25	4566015	3057550	3048289	4579192	0			
23	15251046	11	37	5.56E+08	36.42823	37	37	37	0	37	37	2,11,25	4556463	3048458	3071548	4574577	0			
24	15251046	11	37	5.56E+08	36.42757	37	37	37	0	37	37	2,11,25	4522623	3091054	3074643	4562726	0			
25	15251046	11	37	5.55E+08	36.41759	37	37	37	0	37	37	2,11,25	4545859	3073180	3062739	4569268	0			
26	15251046	11	37	5.55E+08	36.39802	37	37	37	0	37	37	2,11,25	4539780	3066401	3082740	4562125	0			
27	15251046	11	37	5.55E+08	36.39028	37	37	37	0	37	37	2,11,25	4509135	3100679	3082540	4558692	0			
28	15251046	11	37	5.55E+08	36.38156	37	37	37	0	37	37	2,11,25	4535582	3081474	3078151	4555839	0			
29	15251046	11	37	5.55E+08	36.38596	37	37	37	0	37	37	2,11,25	4529426	3068627	3097717	4552576	0			
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31	15251046	2	37	5.55E+08	36.3639	37	37	37	0	37	37	2,11,25	4534267	3088559	3084114	4544105	1			
32	15251046	11	37	5.55E+08	36.37126	37	37	37	0	37	37	2,11,25	4531059	3071760	3104678	4543549	0			
33	15251046	2	37	5.55E+08	36.37091	37	37	37	0	37	37	2,11,25	4500811	3115265	3099633	4535317	20			
34	15251046	11	37	5.55E+08	36.36997	37	37	37	0	37	37	2,11,25	4530975	3092724	3091649	4535701	0			
35	15251046	11	37	5.54E+08	36.34873	37	37	37	0	37	37	2,11,25	4526982	3082707	3106181	4535176	0			
36	15251046	11	37	5.54E+08	36.32999	37	37	37	0	37	37	2,11,25	4502951	3118574	3097842	4531679	0			
37	15251046	2	37	5.54E+08	36.34403	37	37	37	0	37	37	2,11,25	4533745	3092854	3094079	4530349	19			
38	15251046	2	37	5.54E+08	36.33422	37	37	37	0	37	37	2,11,25	4524635	3084990	3112092	4529319	10			
39	15251046	11	37	5.54E+08	36.32853	37	37	37	0	37	37	2,11,25	4501895	3119177	3102552	4527422	0			
40	15251046	2	37	5.54E+08	36.32941	37	37	37	0	37	37	2,11,25	4534150	3096160	3096681	4523629	426			
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46	15251046	2	37	5.54E+08	36.30919	37	37	37	0	37	37	2,11,25	4523672	3105444	3098562	4523236	132			
47	15251046	2	37	5.54E+08	36.30978	37	37	37	0	37	37	2,11,25	4517565	3089757	3118286	4525322	116			
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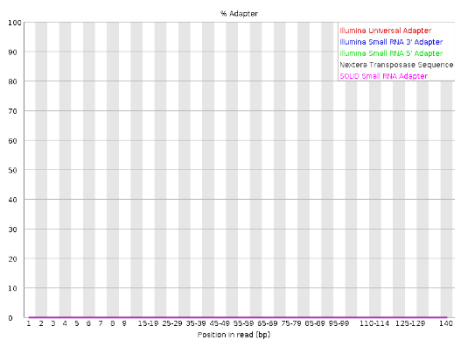
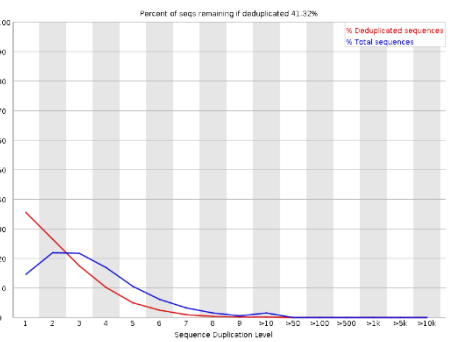
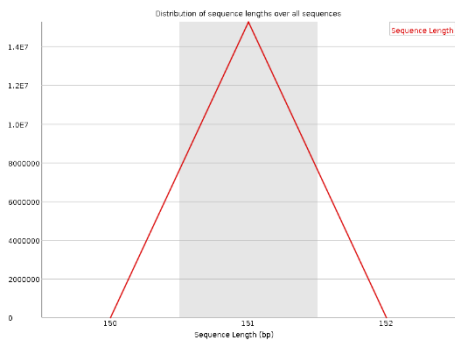
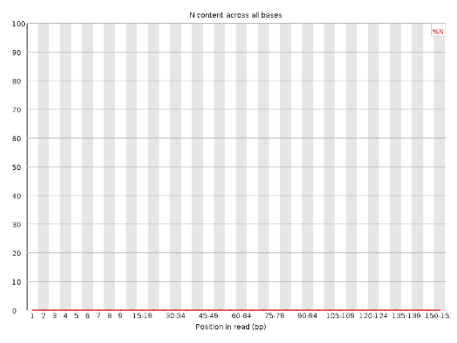
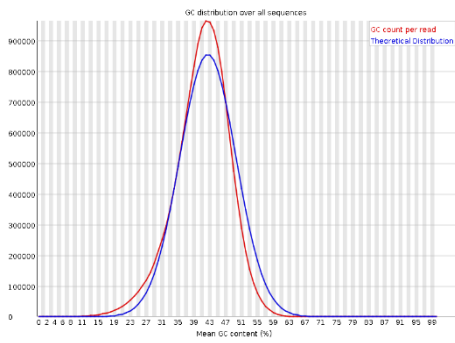
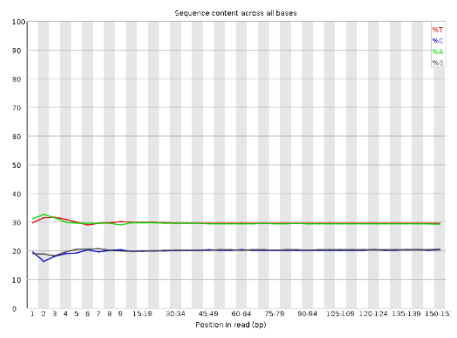
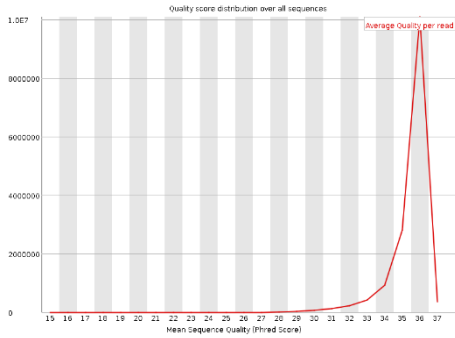
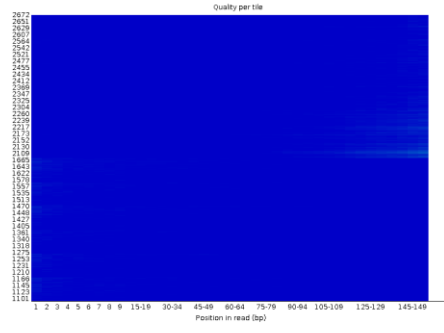
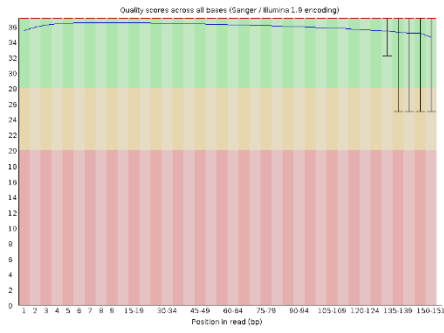
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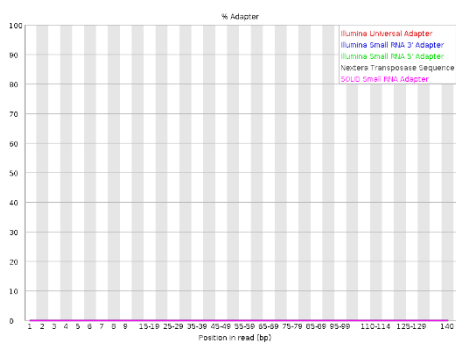
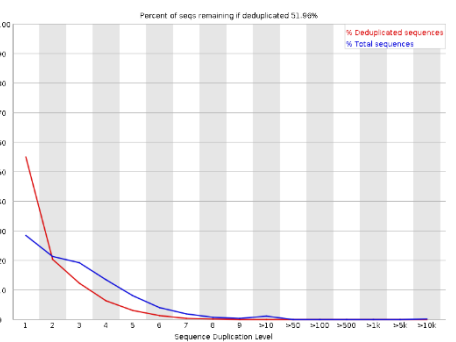
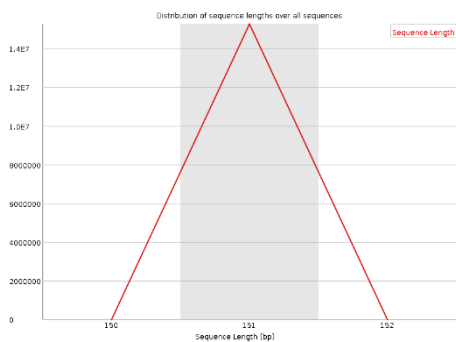
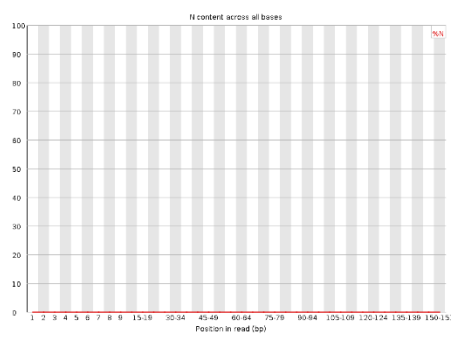
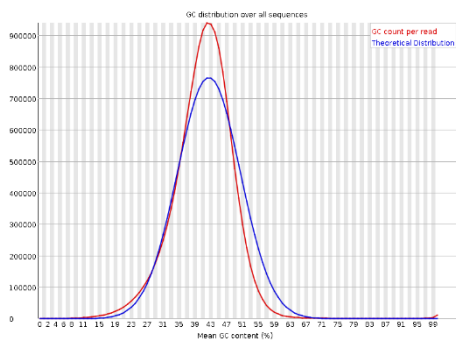
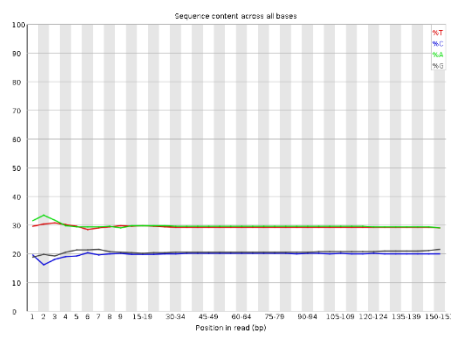
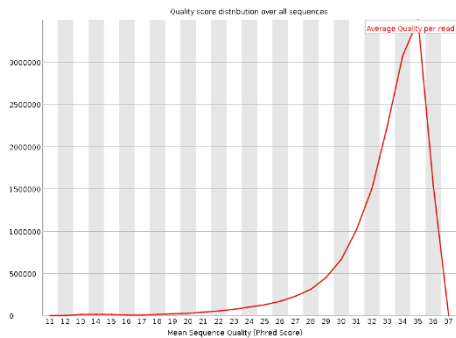
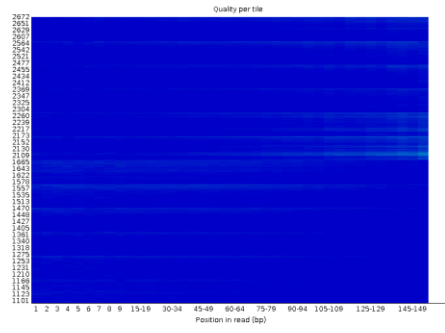
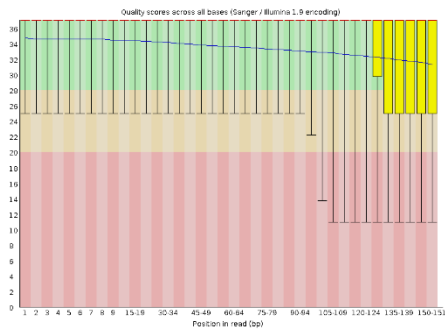
2. Quality assessment of raw reads





From the above graphs for the evolved strain, we can see that while the sequence quality per base remains predominantly constant, the quality score per sequence peaks towards the end of the graph and then falls quickly. We can also see that the percentage of duplicates reduces as we move forward.





From the above graphs for the wild type, we can see that while the sequence quality per base starts to decrease, the quality score per sequence peaks towards the end of the graph and falls again. We can also see that the percentage of duplicates reduces significantly as we move forward.

3. Mapping of genome to reference

<https://usegalaxy.org/datasets/bbd44e69cb8906b590145d713b9c50ec/display/?preview=TRUE>

In the table that can be found above, the evolved strain has been mapped to the reference genome. We can see the differences between the reference and alternate genomes for each position which has a difference.

<https://usegalaxy.org/datasets/bbd44e69cb8906b53e905a98c9039547/display/?preview=TRUE>

In the table that can be found above, the wild type has been mapped to the reference genome. We can see the differences between the reference and alternate genomes for each position which has a difference.

4. Marking and removing duplicate reads

<https://usegalaxy.org/datasets/bbd44e69cb8906b5c7cb0d6e86abfd8d/display/?preview=TRUE>

In the table that can be found above, the duplicate reads have been marked and removed for the BAM file, which contains the mapping of the reference genome to the evolved strain.

<https://usegalaxy.org/datasets/bbd44e69cb8906b59969c829e06f97bb/display/?preview=TRUE>

In the table that can be found above, the duplicate reads have been marked and removed for the BAM file, which contains the mapping of the reference genome to the wild type.

5. Variant calling

<https://usegalaxy.org/datasets/bbd44e69cb8906b590145d713b9c50ec/display/?preview=TRUE>

In the table that can be found above, variants were called for the evolved strain with respect to the reference genome after duplicates had been removed. We can see that most of the mutations are single nucleotide polymorphisms.

<https://usegalaxy.org/datasets/bbd44e69cb8906b53e905a98c9039547/display/?preview=TRUE>

In the table that can be found above, variants were called for the wild type with respect to the reference genome after duplicates had been removed. We can see that most of the mutations are single nucleotide polymorphisms.

6. Annotating variants

<https://usegalaxy.org/datasets/bbd44e69cb8906b50943750f2d471b9a/display/?preview=True>

In the list of tables and illustrations that can be found above, variants were annotated for the evolved strain with respect to the reference genome after variant calling. The 104 mutations have been summarised, and various graphs have been plotted. The ratio of transitions to transversions is found to be 3.2. The length of insertions and deletions has also been found.

<https://usegalaxy.org/datasets/bbd44e69cb8906b5b575f55c1a2ffcca/display/?preview=True>

In the list of tables and illustrations that can be found above, variants were annotated for the wild type with respect to the reference genome after variant calling. The 37 mutations have been summarised, and various graphs have been plotted. The ratio of transitions to transversions is found to be 8, a significant increase from the previous case. The length of insertions and deletions has also been found.

7. Compiling mutations

Wild type

Pos	Ref	Alt	Type of Mutation		Type	Gene	Product
592336	T	G	Substitution	Transversion	CDS	-	putative glycosyltransferase
592340	C	G,T	Substitution		CDS	-	putative glycosyltransferase
592343	G	T	Substitution	Transversion	CDS	-	putative glycosyltransferase
1179108	C	T	Substitution	Transition	CDS	phrB	deoxyribodipyrimidine photolyase (photoreactivation), FAD-binding
1636002	C	A	Substitution	Transversion	CDS	-	putative surface protein (partial adhesin)
1803843	A	G	Substitution	Transition	CDS	-	hypothetical protein; putative membrane protein
1803858	CGCT	TGCC	MNP		CDS	-	hypothetical protein; putative membrane protein
1803900	CTAT	TTAC	MNP		CDS	-	hypothetical protein; putative membrane protein
1803915	G	A	Substitution	Transition	CDS	-	hypothetical protein; putative membrane protein
1803921	A	G	Substitution	Transition	CDS	-	hypothetical protein; putative membrane protein
1803930	TTTA	CTTG	MNP		CDS	-	hypothetical protein; putative membrane protein
1803939	C	T	Substitution	Transition	CDS	-	hypothetical protein; putative membrane protein
1851676	T	G	Substitution	Transversion	CDS	-	uncharacterized prophage protein
2216887	C	A	Substitution	Transversion	CDS	-	putative outer membrane porin protein precursor
2372248	C	G	Substitution	Transversion	CDS	-	conserved hypothetical protein; putative chaperone
2420277	ACCACG	ACCCACG	Insertion		CDS	glnA	glutamine synthetase
2542652	G	A	Substitution	Transition	CDS	lepA	GTP-binding protein
2765010	A	C	Substitution	Transversion	CDS	-	putative periplasmic binding protein of transport/transglycosylase
2803531	G	A	Substitution	Transition	CDS	bap	biofilm associated protein
2804110	G	A	Substitution	Transition	CDS	bap	biofilm associated protein
2804983	G	A	Substitution	Transition	CDS	bap	biofilm associated protein
2805028	A	G	Substitution	Transition	CDS	bap	biofilm associated protein
2805037	A	G	Substitution	Transition	CDS	bap	biofilm associated protein
2805054	T	C	Substitution	Transition	CDS	bap	biofilm associated protein
2805067	G	A	Substitution	Transition	CDS	bap	biofilm associated protein
2805145	G	A	Substitution	Transition	CDS	bap	biofilm associated protein
2805196	A	G	Substitution	Transition	CDS	bap	biofilm associated protein
2843851	T	A	Substitution	Transversion	tRNA	-	Gln tRNA
2843852	A	T	Substitution	Transversion	tRNA	-	Gln tRNA
2844054	CAAAA	GATAT	MNP		tRNA	-	Gln tRNA
3357842	G	A	Substitution	Transition	CDS	-	putative DUF4357 domain-containing protein
3449576	C	A	Substitution	Transversion	CDS	-	conserved hypothetical protein
3449577	T	A	Substitution	Transversion	CDS	-	conserved hypothetical protein
3533222	C	A	Substitution	Transversion	CDS	-	conserved hypothetical protein; putative membrane protein
3533224	AG	AT	Substitution	Transition	CDS	-	conserved hypothetical protein; putative membrane protein
3538607	G	A	Substitution	Transition	CDS	ppc	phosphoenolpyruvate carboxylase

Evolved Strain

Pos	Ref	Alt	Type of Mutation		Type	Gene	Product
18404	G	T	Substitution	Transversion	rRNA	-	16S
82911	GA	TC	MNP		CDS	-	putative UDP-glucose/GDP-mannose dehydrogenase
82918	CAACGTT	GATCCTA	MNP		CDS	-	putative UDP-glucose/GDP-mannose dehydrogenase
173224	T	A	Substitution	Transversion	CDS	-	conserved hypothetical protein; putative dyp-type peroxidase
173228	GCATA	TGATC	MNP		CDS	-	conserved hypothetical protein; putative dyp-type peroxidase
173233	A	C	Substitution	Transversion	CDS	-	conserved hypothetical protein; putative dyp-type peroxidase
178458	AGGGGGGGGGCTCT	AGGGGGGGGGCTCT	Deletion		CDS	atpI	ATP synthase protein I
449101	CAACGTGT	CGTAGATGA	Insertion		CDS	-	putative acyl-CoA dehydrogenase (AidB)
449276	TAT	CAA	MNP		CDS	-	putative acyl-CoA dehydrogenase (AidB)
449279	TTTTCAGCATA	ATTTAGAATC,TTTTAGAATC	MNP		CDS	-	putative acyl-CoA dehydrogenase (AidB)
592336	T	G	Substitution	Transversion	CDS	-	putative glycosyltransferase
592338	TTC	TTG			CDS	-	putative glycosyltransferase
786523	A	T	Substitution	Transversion	tRNA	-	Asp tRNA
935746	AGTCA	TGATC			CDS	-	conserved hypothetical protein
941297	TCATAAA	AAATATAAG	Insertion		IS	IS1236	Insertion Sequence IS1236, IS3 family
941305	G	A	Substitution	Transition	IS	IS1236	Insertion Sequence IS1236, IS3 family
942543	AAGTGATAAT	AAGGATTGATATT	Insertion		CDS	-	conserved hypothetical protein
942553	TA	TT			CDS	-	conserved hypothetical protein
942556	A	G	Substitution	Transition	CDS	-	conserved hypothetical protein
943612	G	A	Substitution	Transition	CDS	-	conserved hypothetical protein
943613	A	T	Substitution	Transversion	CDS	-	conserved hypothetical protein
943615	AATTAT	AAAGAT	MNP		CDS	-	conserved hypothetical protein
944841	TATTTGG	GATACTGT	MNP		IS	IS1236	Insertion Sequence IS1236, IS3 family
944857	AATTG	ATAATC,ATCTTT,AGATGG,AATCTA	Insertion		CDS	-	conserved hypothetical protein; putative membrane protein
944863	C	G	Substitution	Transversion	CDS	-	conserved hypothetical protein; putative membrane protein
944865	GC	AG,GT	MNP		CDS	-	conserved hypothetical protein; putative membrane protein
944867	TT	TC	Substitution	Transition	CDS	-	conserved hypothetical protein; putative membrane protein
964810	CATCAGG	CTGATCCTA	Insertion		CDS	vanR	transcriptional regulator for ferulate or vanillate catabolism (GntR family)
968302	TGGATA	GGCAATTA	Insertion		CDS	-	hypothetical protein
968310	GATATG	CATAAT	MNP		CDS	-	hypothetical protein
1168348	TTGT	GATC	MNP		CDS	-	conserved hypothetical protein
1168357	TCG	GAT	MNP		CDS	-	conserved hypothetical protein
1179108	C	T	Substitution	Transition	CDS	phrB	deoxyribodipyrimidine photolyase (photoreactivation), FAD-binding
1202039	G	A	Substitution	Transition	CDS	hemL	glutamate-1-semialdehyde aminotransferase
1360293	TGGCT	TCT	Deletion		CDS	clpA	ATP-binding protease component
1501968	T	G	Substitution	Transversion	CDS	-	putative Dibenzothiophene desulfurization enzyme
1501969	GGTCC	GATCA	MNP		CDS	-	putative Dibenzothiophene desulfurization enzyme
1501978	TTAT	GATC	MNP		CDS	-	putative Dibenzothiophene desulfurization enzyme
1522654	TCATAT	TAGGATCA	Insertion		CDS	-	putative transport protein (ABC superfamily, ATP_bind)
1522664	ACG	GAT	MNP		CDS	-	putative transport protein (ABC superfamily, ATP_bind)
1534825	T	G	Substitution	Transversion	CDS	-	putative tonB-dependent receptor protein (outer membrane salicin receptor)
1534829	ACTTTT	AATGC	Deletion		CDS	-	putative tonB-dependent receptor protein (outer membrane salicin receptor)
1572873	TTTTG	GTCTA	MNP		CDS	-	putative transcriptional regulator (Tetr family)
1572878	GGATCG	GGATCACG	Insertion		CDS	-	putative transcriptional regulator (Tetr family)
1636002	C	A	Substitution	Transversion	CDS	-	putative surface protein (partial adhesion)
1708197	G	A	Substitution	Transition	CDS	pcaU	regulatory protein for pca operon (activator)
1723516	CAGCAG	GATCCTA,CAGCTG	Insertion		CDS	quiA	quininate/shikimate dehydrogenase [Pyroloquinoline-quinone] (NAD(P)-independent quininate dehydrogenase)
1729396	AGAAGA	TGATCC			CDS	hcaE	porin
1729596	ATTGGT	CTAGGATC	Insertion		CDS	hcaE	porin
1789246	ACAT	GATC	MNP		CDS	ppk	polyphosphate kinase (Polyphosphoric acid kinase) (ATP-polyphosphate phosphotransferase)
1789408	TTTATT	GCTAGG	MNP		CDS	ppk	polyphosphate kinase (Polyphosphoric acid kinase) (ATP-polyphosphate phosphotransferase)
1803858	CGCT	TGCC	MNP		CDS	-	hypothetical protein; putative membrane protein
1803900	CTAT	TTAC	MNP		CDS	-	hypothetical protein; putative membrane protein

1803915	G	A	Substitution	Transition	CDS	-	hypothetical protein; putative membrane protein
1803921	A	G	Substitution	Transition	CDS	-	hypothetical protein; putative membrane protein
1803930	TTTA	CTTG	MNP		CDS	-	hypothetical protein; putative membrane protein
1803939	C	T	Substitution	Transition	CDS	-	hypothetical protein; putative membrane protein
1845223	TGCGG	GGATC	MNP		CDS	-	putative transcriptional regulator (AraC family)
1845232	AA	TG	MNP		CDS	-	putative transcriptional regulator (AraC family)
1845236	A	C	Substitution	Transversion	CDS	-	putative transcriptional regulator (AraC family)
1857795	A	C	Substitution	Transversion	CDS	-	putative repressor of phage gene expression protein (RstR-like)
1858199	C	G	Substitution	Transversion	CDS	-	conserved hypothetical protein
2164696	ATATAAGGTTATTGA	GGATCAGTTATTGA,ATATAAGGTGATCCT	Deletion/MNP		CDS	-	conserved hypothetical protein
2216887	C	A	Substitution	Transversion	CDS	-	putative outer membrane porin protein precursor
2372312	G	T	Substitution	Transversion	CDS	-	conserved hypothetical protein; putative chaperone
2420277	ACCACG	ACCCACG	Insertion		CDS	glnA	glutamine synthetase
2448584	AG	TC	MNP		CDS	-	putative two-component response regulator protein
2448591	AACGG	GATCC	MNP		CDS	-	putative two-component response regulator protein
2542652	G	A	Substitution	Transition	CDS	lepA	GTP-binding protein
2667494	T	G	Substitution	Transversion	fCDS	-	fragment of component of DNA polymerase V (UmuC) (part 1)
2753757	TC	CA	MNP		CDS	-	conserved hypothetical protein
2753764	TTTT	ATCC	MNP		CDS	-	conserved hypothetical protein
2754220	CGCATT	AGGATC	MNP		CDS	-	putative transcriptional regulator (lclR family)
2754229	CTAGA	CAGA	Deletion		CDS	-	putative transcriptional regulator (lclR family)
2754235	C	T	Substitution	Transition	CDS	-	putative transcriptional regulator (lclR family)
2754236	T	A	Substitution	Transversion	CDS	-	putative transcriptional regulator (lclR family)
2754238	A	G	Substitution	Transition	CDS	-	putative transcriptional regulator (lclR family)
2765010	A	C	Substitution	Transversion	CDS	-	putative periplasmic binding protein of transport/transglycosylase
2803531	G	A	Substitution	Transition	CDS	bap	biofilm associated protein
2804110	G	A	Substitution	Transition	CDS	bap	biofilm associated protein
2804938	A	G	Substitution	Transition	CDS	bap	biofilm associated protein
2804956	T	C	Substitution	Transition	CDS	bap	biofilm associated protein
2804983	G	A	Substitution	Transition	CDS	bap	biofilm associated protein
2805028	A	G	Substitution	Transition	CDS	bap	biofilm associated protein
2805037	A	G	Substitution	Transition	CDS	bap	biofilm associated protein
2805054	T	C	Substitution	Transition	CDS	bap	biofilm associated protein
2805067	G	A	Substitution	Transition	CDS	bap	biofilm associated protein
2805145	G	A	Substitution	Transition	CDS	bap	biofilm associated protein
2805196	A	G	Substitution	Transition	CDS	bap	biofilm associated protein
2843816	G	C	Substitution	Transversion	tRNA	-	Gln tRNA
2843851	T	A	Substitution	Transversion	tRNA	-	Gln tRNA
2843852	A	T	Substitution	Transversion	tRNA	-	Gln tRNA
2843853	C	A	Substitution	Transversion	tRNA	-	Gln tRNA
2843856	C	A	Substitution	Transversion	tRNA	-	Gln tRNA
2843964	G	T	Substitution	Transversion	tRNA	-	Gln tRNA
2844056	A	T	Substitution	Transversion	tRNA	-	Gln tRNA
2937249	A	C	Substitution	Transversion	CDS	ksgA	S-adenosylmethionine-6-N',N'-adenosyl (rRNA) dimethyltransferase, kasugamycin resistance
2937252	A	C	Substitution	Transversion	CDS	ksgA	S-adenosylmethionine-6-N',N'-adenosyl (rRNA) dimethyltransferase, kasugamycin resistance
2937253	A	C	Substitution	Transversion	CDS	ksgA	S-adenosylmethionine-6-N',N'-adenosyl (rRNA) dimethyltransferase, kasugamycin resistance
3241277	G	A	Substitution	Transition	CDS	comP	competence factor involved in DNA binding and uptake
3241284	C	T	Substitution	Transition	CDS	comP	competence factor involved in DNA binding and uptake
3241285	AA	CC,TA			CDS	comP	competence factor involved in DNA binding and uptake
3241287	T	G	Substitution	Transversion	CDS	comP	competence factor involved in DNA binding and uptake
3329806	TATTG	GATCA	MNP		CDS	-	putative outer membrane protein precursor
3329814	AAG	TGA	MNP		CDS	-	putative outer membrane protein precursor
3357842	G	A	Substitution	Transition	CDS	-	putative outer membrane protein precursor

Learning Experience

As a chemical engineering student with little prior exposure to biotechnology, this internship and the processes that led up to the task gave me something new to learn.

I learnt about a new way to be able to make greener fuel with this study, something I had not known before. This was a unique learning experience.

I began by making a literature study where I learnt about how actually to perform a literature study. I read several papers about whole-genome sequence analysis and adaptive laboratory evolution, which taught me what these processes were and how they could be implemented. I learnt about the applications of these concepts and gave myself an introduction to the work I had to perform.

I learnt about different bioinformatics tools that can be used to perform various tasks in the pipeline for whole-genome sequencing. I learnt about what kind of codes these tools use, how they work, and what results they produce.

Subsequently, I learnt about mutations, their types, and their effects. I was able to use this knowledge when compiling the final spreadsheet.

This experience gave me an exciting introduction to this branch of science and gave me even more motivation to pursue biotechnology further. I am incredibly grateful for the people involved, who taught me with patience and understanding.

Conclusion

Climate change has become a growing concern that needs to be given serious consideration. Today's population and their demands result in the use of petroleum products which are non-renewable, non-eco-friendly sources of energy. This study takes us one step closer to realising the goal of production of renewable and harmless sources of energy.

A single mutation can have a significant effect, but in many cases, evolutionary change is based on the accumulation of many mutations with minor consequences. Mutational effects can be beneficial, harmful, or neutral, depending on their context or location. Most non-neutral mutations are deleterious. In general, the more base pairs that are affected by a mutation, the more significant is the effect of the mutation, and the larger is the mutation's probability of being deleterious.

Biofilm associated protein (BAP) [\[10\]](#) plays a vital role in biofilm formation and adhesion to host cells in *Acinetobacter*. A possible outcome is that the mutation of a *bap* gene of a coding sequence could result in the sequence of proteins changing. A likely outcome is a change in film properties and adhesive properties of the strain. This can be used to enhance the breakdown of lignin.

The mutations of tRNA [\[11\]](#) affect how the ribosomes read the mRNA. This affects the fidelity of the decoding process. The tRNA adopts a different conformation when it reaches a ribosome that correctly decodes it. This could affect the protein structure, which could result in a change in the conformation of these proteins.

Thus, the whole-genome sequence analysis of *Acinetobacter baylyi* has been performed using computational methods as explained previously. The predictions made have to be validated by conducting experiments. When the pattern of mutations and their effects are studied, the strain can be modified further to produce desirable outcomes. These mutations in the evolved strain would be used to uptake the lignin monomers and produce a fuel that can be used in place of petroleum products without damaging the environment.

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