

Anti-ds-DNA Screen (A,G, M) ELISA

For the quantitative and qualitative determination of IgA/G/M antibodies against doublestranded DNA (dsDNA) in human serum

For Research Use Only. Not For Use In Diagnostic Procedures.

Catalog Number: 35-DSSHU-E01

Size: 96 wells

Version: 003: 2015-10-28 - ALPCO 1.1

1. Intended Use

Anti-ds-DNA Screen (IgA, IgG, IgM) ELISA is a solid phase enzyme immunoassay with human recombinant double-stranded DNA (dsDNA) for the quantitative and qualitative detection of IgG, IgA, and IgM antibodies against dsDNA in human serum.

Anti-dsDNA antibodies mainly recognize the phosphate units of DNA, thus these autoantibodies also bind single stranded DNA (ssDNA). To ensure correct quantitation of anti-dsDNA antibodies the antigen used in the kit has been proven to be free of contamination with ssDNA.

For Research Use Only. Not For Use In Diagnostic Procedures.

2. Application and Principle of the Assay

Antibodies binding to DNA belong to the group of anti-nuclear antibodies (ANA) that have been observed in several autoimmune diseases. Antibodies reacting with native double-stranded (ds) DNA are regarded as being specific for systemic lupus erythematosus (SLE) and have been observed in approximately 50-80% of the individuals with SLE.

Antibodies against dsDNA are found during active phases of SLE. The serum concentration is positively correlated with the severity of the disease. Thus, detection of these autoantibodies is important in the study of SLE.

Most individuals with SLE display IgG class antibodies against dsDNA. These autoantibodies are associated with lupus nephritis. In addition, approximately 30% of individuals with SLE develop IgA class anti-dsDNA antibodies. It has been suggested that the presence of these IgA class anti-dsDNA antibodies may define a certain subset of individuals with SLE. Studies demonstrated the association of this subclass with certain parameters of the disease activity, such as elevated erythrocyte sedimentation rate, or the consumption of complement component C3, as well as the clinical parameters of cutaneous vasculitis, acral necrosis, and erythema. No association was found for nephritis and arthritis.

IgM class anti-dsDNA antibodies were found in 52% of the sera from individuals with SLE. In contrast to IgG and IgA class autoantibodies, the subclass IgM antibodies do not correlate with disease activity. However, a highly significant negative correlation between IgM anti-dsDNA antibodies and lupus nephritis, including its laboratory parameters, was demonstrated. Therefore, IgM class anti-dsDNA antibodies may indicate a subset of individuals with lupus being protected against the risk of developing nephritis.

Principle of the test

Serum samples diluted 1:101 are incubated in the microplates coated with the specific antigen. Antibodies, if present in the sample, bind to the antigen. The unbound fraction is washed off in the following step. Afterwards, anti-human immunoglobulins conjugated to horseradish peroxidase (conjugate) are incubated and react with the antigen-antibody complex of the samples in the microplates. Unbound conjugate is washed off in the following step. Addition of TMB substrate generates an enzymatic colorimetric (blue) reaction, which is stopped by diluted acid (color changes to yellow). The intensity of color formation from the chromogen is a function of the

amount of conjugate bound to the antigen-antibody complex and this is proportional to the initial concentration of the respective antibodies in the sample.

3. Kit Contents

| TO BE RECONSTITUTED | | | | |
|---------------------|-----------------------|--------------|----------------|--|
| Item | Quantity | Cap color | Solution color | Description / Contents |
| Sample Buffer (5x) | 1 x 20mL | White | Yellow | 5x concentrated Tris, sodium chloride (NaCl), bovine serum albumin (BSA), sodium azide < 0.1% (preservative) |
| Wash Buffer (50x) | 1 X 20mL | White | Green | 50x concentrated Tris, NaCl, Tween 20, sodium azide < 0.1% (preservative) |
| | | RE | ADY TO USE | |
| Item | Quantity | Cap color | Solution color | Description / Contents |
| Negative Control | 1 x 1.5mL | Green | Colorless | Human serum (diluted), bovine serum albumin (BSA), sodium azide < 0.1% (preservative) |
| Positive Control | 1 x 1.5mL | Red | Yellow | Human serum (diluted), bovine serum albumin (BSA), sodium azide < 0.1% (preservative) |
| Cut-off Calibrator | 1 x 1.5mL | Blue | Yellow | Human serum (diluted), bovine serum albumin (BSA), sodium azide < 0.1% (preservative) |
| Calibrators | 6 x 1.5mL | White | Yellow * | Concentration of each calibrator: 0, 3, 10, 30, 100, 300 U/mL. Human serum (diluted), bovine serum albumin (BSA), sodium azide < 0.1% (preservative) |
| Conjugate, IgA/G/M | 1 x 15mL | White | Red | Containing: Anti-human immunoglobulins conjugated to horseradish peroxidase, bovine serum albumin (BSA) |
| TMB Substrate | 1 x 15mL | Black | Colorless | Stabilized tetramethylbenzidine and hydrogen peroxide (TMB/H ₂ O ₂) |
| Stop Solution | 1 x 15mL | White | Colorless | 1M Hydrochloric Acid |
| Microtiter plate | 12 x 8 well strips | N/A | N/A | With breakaway microwells. Refer to paragraph 1 for coating. |

^{*} Color increasing with concentration

MATERIALS REQUIRED, BUT NOT PROVIDED

Microtiter plate reader 450 nm reading filter and recommended 620 nm reference filter (600-690 nm). Glassware (cylinder 100-1000mL), test tubes for dilutions. Vortex mixer, precision pipettes (10, 100, 200, 500, 1000 μ L) or adjustable multipipette (100-1000 μ L). Microplate washing device (300 μ L repeating or multichannel pipette or automated system), adsorbent paper. Our tests are designed to be used with purified water according to the definition of the United States Pharmacopeia (USP 26 - NF 21) and the European Pharmacopeia (Eur.Ph. 4th ed.).

4. Storage and Shelf Life

Store all reagents and the microplate at 2-8°C/35-46°F, in their original containers. Once prepared, reconstituted solutions are stable for 1 month at 2-8°C/35-46°F. Reagents and the microplate shall be used within the expiry date indicated on each component, only. Avoid intense exposure of TMB solution to light. Store microplates in designated foil, including the desiccant, and seal tightly.

5. Precautions of Use

5.1. Health Hazard Data

THIS PRODUCT IS FOR RESEARCH USE ONLY. Although this product is not considered particularly toxic or dangerous in conditions of the intended use, refer to the following for maximum safety:

Recommendations and precautions

This kit contains potentially hazardous components. Though kit reagents are not classified being irritant to eyes and skin, it is recommended to avoid contact with eyes and skin and wear disposable gloves.

WARNING! Calibrators, Controls and Buffers contain sodium azide (NaN₃) as a preservative. NaN₃ may be toxic if ingested or adsorbed by skin or eyes. NaN₃ may react with lead and copper plumbing to form highly explosive metal azides. On disposal, flush with a large volume of water to prevent azide build-up. Please refer to decontamination procedures as outlined by CDC or other local/national guidelines.

Do not smoke, eat or drink when manipulating the kit. Do not pipette by mouth.

All human source material used for some reagents of this kit (e.g. controls, standards) has been tested by approved methods and found negative for HbsAg, Hepatitis C and HIV 1. However, no test can guarantee the absence of viral agents in such material completely. Thus, handle kit controls, standards and samples as if capable of transmitting infectious diseases and according to national requirements.

The kit contains material of animal origin as stated in the table of contents, handle according to national requirements.

5.2. General Directions for Use

In case that the product information, including the labeling, is defective or incorrect please contact ALPCO.

Do not mix or substitute Controls, Calibrators, Conjugates or microplates from different lot numbers. This may lead to variations in the results.

Allow all components to reach room temperature (20-32°C/68-89.6°F) before use, mix well and follow the recommended incubation scheme for an optimum performance of the test.

Incubation: It is recommended to test performance at 30°C/86°F for automated systems.

Never expose components to a higher temperature than 37°C/98.6°F.

Always pipette substrate solution with brand new tips only. Protect this reagent from light. Never pipette conjugate with tips used with other reagents prior.

6. Sample Collection, Handling, and Storage

Use preferentially freshly collected serum samples. Blood withdrawal must follow national requirements. Do not use icteric, lipemic, hemolyzed or bacterially contaminated samples. Sera with particles should be cleared by low speed centrifugation (<1000 xg).

After separation, the serum samples should be used during the first 8 hours, respectively stored tightly closed at 2-8°C/35-46°F up to 48 hours, or frozen at -20°C/-4°F for longer periods.

7. Assay Procedure

7.1. Preparations Prior to Starting

Dilute concentrated reagents:

Dilute the concentrated sample buffer 1:5 with distilled water (e.g. 20 mL plus 80 mL).

Dilute the concentrated wash buffer 1:50 with distilled water (e.g. 20 mL plus 980 mL).

To avoid mistakes we suggest to mark the cap of the different calibrators.

Samples:

Dilute serum samples 1:101 with sample buffer (1x)

e.g. 1000 µL sample buffer (1x) + 10 µL serum. Mix well!

Washing:

Prepare 20 mL of diluted wash buffer (1x) per 8 wells or 200 mL for 96 wells

e.g. 4 mL concentrate plus 196 mL distilled water

Automated washing:

Consider excess volumes required for setting up the instrument and dead volume of robot pipette.

Manual washing:

Discard liquid from wells by inverting the plate. Knock the microwell frame with wells downside vigorously on clean adsorbent paper. Pipette 300 μ L of diluted wash buffer into each well, wait for 20 seconds. Repeat the whole procedure twice again.

Microplates:

Calculate the number of wells required for the test. Remove unused wells from the frame, replace and store in the provided plastic bag (2-8°C/35-46°F), together with desiccant and seal tightly.

7.2. Assay Procedure

Ensure preparation from step 7.1 above have been carried out prior to pipetting.

Use the following steps in accordance with quantitative or qualitative interpretation:

| CalA: Calibrator A | CalD: Calibrator A | PC: Positive Control | S1: Sample 1 |
|--------------------|--------------------|------------------------|--------------|
| CalB: Calibrator B | CalE: Calibrator A | NC: Negative Control | S2: Sample 2 |
| CalC: Calibrator C | CalF: Calibrator A | CC: Cut-off Calibrator | S3: Sample 3 |

| For Quantitative Interpretation | | | | |
|---------------------------------|-------|-------|------------|---|
| | 1 | 2 | 3 | 4 |
| Α | Cal A | Cal E | <i>S</i> 1 | |
| В | Cal A | Cal E | S1 | |
| С | Cal B | Cal F | S2 | |
| D | Cal B | Cal F | <i>S2</i> | |
| E | Cal C | PC | <i>S3</i> | |
| F | Cal C | PC | <i>S3</i> | |
| G | Cal D | NC | ••• | |
| Н | Cal D | NC | | |

| For Qualitative Interpretation | | | | |
|--------------------------------|----|-----------|---|---|
| | 1 | 2 | 3 | 4 |
| Α | СС | 52 | | |
| В | СС | 52 | | |
| C | PC | <i>S3</i> | | |
| D | PC | S3 | | |
| Ε | NC | | | |
| F | NC | | | |
| G | S1 | | | |
| Н | S1 | ••• | | |

- 1. Pipette 100 µL into the designated wells either:
 - a. calibrators (CalA to CalF) for quantitative interpretation **OR**
 - b. cut-off calibrator for qualitative interpretation
- 2. Pipette 100µL of each of the following into the designated wells:
 - a. Positive Control (PC)
 - b. Negative Control (NC)
- 3. Pipette 100 µL of each diluted serum sample into the designated microwells.
- 4. Incubate for 30 minutes at 20-32°C/68-89.6°F.
- 5. Wash 3x with 300 µL wash buffer (diluted 1:50).
- 6. Pipette 100 μL conjugate into each well.
- 7. Incubate for 30 minutes at 20-32°C/68-89.6°F.
- 8. Wash 3x with 300 µL wash buffer (diluted 1:50).
- 9. Pipette 100 µL TMB substrate into each well.
- 10. Incubate for 30 minutes at 20-32°C/68-89.6°F, protected from intense light.
- 11. Pipette 100 µL stop solution into each well, using the same order as pipetting the substrate.
- 12. Incubate for 5 minutes minimum.
- 13. Agitate plate carefully for 5 sec.
- 14. Read absorbance at 450 nm (optionally 450/620 nm) within 30 minutes.

8. Quantitative and Qualitative Interpretation

For **quantitative interpretation** establish the standard curve by plotting the optical density (OD) of each calibrator (y-axis) with respect to the corresponding concentration values in U/mL (x-axis). For best results it is recommended to use log/lin coordinates and 4-Parameter Fit. From the OD of each sample, read the corresponding antibody concentrations expressed in **U/mL**.

| Equivocal Range | Positive Results |
|-----------------|------------------|
| 16 - 24 U/mL | >24 U/mL |

Example of a standard curve

It is recommended to pipette calibrators in parallel for each run.

| Calibrators IgA/G/M | OD 450/620 nm | CV % (Variation) |
|---------------------|---------------|------------------|
| 0 U/mL | 0.036 | 2.9 |
| 3 U/mL | 0.176 | 2.3 |
| 10 U/mL | 0.314 | 2.9 |
| 30 U/mL | 0.618 | 2.9 |
| 100 U/mL | 1.312 | 0.1 |
| 300 U/mL | 2.076 | 0.7 |

Example of calculation

| Sample | Replicate (OD) | Mean (OD) | Result (U/mL) |
|--------|-------------------|--------------|------------------|
| S1 | 0.799/0.744 | 0.772 | 40.3 |
| S2 | 1.404/1.393 | 1.399 | 119.5 |

Samples above the calibrator range should be reported as >Max. They should be diluted as appropriate and re-assayed. Samples below calibrator range should be reported as <Min.

For lot specific data, see enclosed quality control leaflet. In house quality controls and/or internal pooled sera, as foreseen by national regulations or laboratory protocols may be used.

Each laboratory should establish its own normal range based upon its own techniques, controls, equipment, and sample population according to their own procedures.

In case that the values of the controls do not meet criteria, the assay is invalid and has to be repeated.

The following technical issues should be verified: expiration date of (prepared) reagents, storage conditions, pipettes, devices, photometer, incubation conditions, and washing methods.

If the items tested show aberrant values or any kind of deviation or that the validation criteria are not met without explicable cause, please contact ALPCO.

For **qualitative interpretation** read the optical density of the cut-off calibrator and the samples. Compare sample OD values with the OD of the cut-off calibrator. For qualitative interpretation, it is recommended to consider sera within a range of 20% around the cut-off value as equivocal. All samples with higher ODs are considered positive, samples with lower ODs are considered negative.

Negative: OD sample < 0.8 x OD cut-off

Equivocal: $0.8 \times OD \text{ cut-off } \leq OD \text{ sample } \leq 1.2 \times OD \text{ cut-off}$

Positive: OD sample > 1.2 x OD cut-off

9. Technical Data

Sample material: serum

Sample volume: 10 µL of sample diluted 1:101 with 1x sample buffer

Total incubation time: 90 minutes at 20-32°C/68-89.6°F

Calibration range: 0-300 U/mL

Analytical sensitivity: 1.0 U/mL

Storage: at 2-8°C/35-46°F use original vials only

Number of determinations: 96 tests

10. Performance Data

10.1. Analytical sensitivity

Testing sample buffer 30 times on the *Anti-ds-DNA Screen (A,G,M) ELISA* gave an analytical sensitivity of 1.0 U/mL.

10.2. Specificity and sensitivity

The microplates are coated with **recombinant human dsDNA**. No cross-reactivities to other autoantigens have been found. Antibodies targeting dsDNA demonstrate a sensitivity of 85% for SLE. Combining all three immunoglobulin subclasses results in a testing sensitivity of 90%.

10.3. Linearity:

Chosen sera have been tested with this kit and found to dilute linearly. However, due to the heterogeneous nature of human autoantibodies there might be samples that do not follow this rule.

| | | Measured | Expected | |
|---------------|--------------------|----------------------|----------------------|-----------------|
| Sample No. | Dilution Factor | Concentration (U/mL) | Concentration (U/mL) | Recovery (%) |
| 1 | 1 / 100 | 42.9 | 43.2 | 99.3 |
| | 1 / 200 | 20.4 | 21.6 | 99.4 |
| | 1 / 400 | 9.3 | 10.8 | 86.1 |
| | 1 / 800 | 4.9 | 5.4 | 90.7 |
| 2 | 1 / 100 | 179.4 | 176.0 | 101.9 |
| | 1 / 200 | 8-6.4 | 88.0 | 98.2 |
| | 1 / 400 | 41.8 | 44.0 | 95.0 |
| | 1 / 800 | 19.8 | 22.0 | 90.0 |

10.4. Precision

To determine the precision of the assay, the variability (intra- and inter-assay) was assessed by examining its reproducibility on three serum samples selected to represent a range over the standard curve.

| Intra-Assay | | | |
|-------------|--------|-----|--|
| Sample | Mean | CV | |
| No. | (U/mL) | (%) | |
| 1 | >300.0 | 2.1 | |
| 2 | 138.0 | 2.4 | |
| 3 | 26.4 | 4.7 | |

| ı | Inter-Assay | | | |
|--------|-------------|-----|--|--|
| Sample | Mean | CV | | |
| No. | (U/mL) | (%) | | |
| 1 | 463.3 | 2.6 | | |
| 2 | 171.6 | 2.3 | | |
| 3 | 58.2 | 4.6 | | |

10.5. Calibration

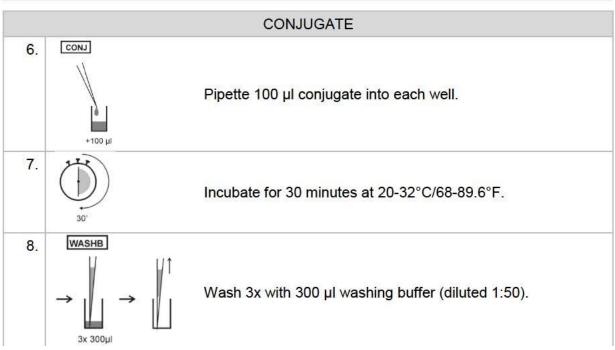
Due to the lack of international reference calibration this assay is calibrated in arbitrary units (U/mL).

11. Literature

- 1. Tan EM, Cohen AS, Fries JF, et al. (1982). Revised criteria for the classification of system lupus erythematosus. Arthritis Rheumatism 25: 1271-1277.
- 2. Witte T, Hartung K, Matthias T, Sachse C, Fricke M, Deicher H, Kalden JR, Lakomek HJ, Peter HH, Schmidt RE (1998). Association of IgA anti-dsDNA antibodies with vasculitis and disease activity in systemic lupus erythematosus. Rheumatol Int 18: 63-69.
- 3. Witte T, Hartung K, Sachse C, Matthias T, Fricke M, Deicher H, Kalden JR, Lakomek HJ, Peter HH, Schmidt RE, SLE study group (1998). IgM anti-dsDNA antibodies in systemic lupus erythematosus: negative association with nephritis. Rheumatol Int 18: 85-91.

Annex A: Test Procedure

| Step | Description |
|------|---|
| 1. | Ensure preparations from step 7.1 above have been carried out prior to pipetting. |
| 2. | Use the following steps in accordance with quantitative/ qualitative interpretation results desired: |
| | CONTROLS & SAMPLES |
| 3. | Pipette into the designated wells as described in chapter 7.2 above, 100 μl of either: a. Calibrators (CAL.A to CAL.F) for <i>QUANTITATIVE</i> or b. Cut-off Calibrator (CC) for <i>QUALITATIVE</i> interp. and 100 μl of each of the following: • Negative control (NC) and Positive control (PC), and |
| 4. | Patients diluted serum (P1, P2) Incubate for 30 minutes at 20-32°C/68-89.6°F. |
| 5. | → WashB WashB Wash 3x with 300 μl washing buffer (diluted 1:50). |



| | SUBSTRATE | | | | |
|-----|--|---|--|--|--|
| 9. | +100 µl | Pipette 100 μl TMB substrate into each well. | | | |
| 10. | 300 | Incubate for 30 minutes at 20-32°C/68-89.6°F, protected from intense light. | | | |
| | | STOP | | | |
| 11. | STOP → +100 μI | Pipette 100 µl stop solution into each well, using the same order as pipetting the substrate. | | | |
| 12. | 5' | Incubate 5 minutes minimum. | | | |
| 13. | | Agitate plate carefully for 5 sec. | | | |
| 14. | OD ₅₅ OD ₆₂₀ 450/620 nm | Read absorbance at 450 nm (recommended 450/620 nm) within 30 minutes. | | | |