Filtration of submicrometer particles by pelagic tunicates

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Salps are common in oceanic waters and have higher per-individual filtration rates than any other zooplankton filter feeder. Although salps are centimeters in length, feeding via particle capture occurs on a fine, mucous mesh (fiber diameter d ~0.1 μm) at low velocity (U = 1.6 ± 0.6 cm·s⁻¹, mean ± SD) and is thus a low Reynolds-number (Re ≈ 10⁻⁵) process. In contrast to the current view that particle encounter is dictated by simple sieving of particles larger than the mesh spacing, a low-Re mathematical model of encounter rates by the salp feeding apparatus for realistic oceanic particle-size distributions shows that submicron particles, due to their higher abundances, are encountered at higher rates (particles per time) than larger particles. Data from feeding experiments with 0.5-, 1-, and 3-μm diameter polystyrene spheres corroborate these findings. Although particles larger than 1 μm (e.g., flagellates, small diatoms) represent a larger carbon pool, smaller particles in the 0.1- to 1-μm range (e.g., bacteria, Prochlorococcus) may be more quickly digestible because they present more surface area, and we find that particles smaller than the mesh size (1.4 μm) can fully satisfy salp energetic needs. Furthermore, by packaging submicrometer particles into rapidly sinking fecal pellets, pelagic tunicates can substantially change particle-size spectra and increase downward fluxes in the ocean.

Filtration rates, small mesh size, and rapid pellet sinking imply that salps have the potential to shift particle distributions toward larger sizes, contribute to vertical transport, and remove substantial amounts of primary production from surface waters. These impacts will be particularly profound following population increases, which can occur suddenly under favorable conditions due to short generation times and a two-part life cycle comprising asexually reproducing individuals and pseudocolonial chains of sexually reproducing salps (1).

Generally, encounter rates between particles and filter elements depend on the Reynolds number (Re = dU/ν, where d is mesh fiber diameter, U is velocity, and ν is kinematic viscosity), which measures the relative importance of inertial and viscous forces. At low Re (Re ≪ 1), viscous effects prevail and prevent flow separation around filter elements (9). Filtration in salps operates in this regime, as Re ≈ 2×10⁻³, based on mesh fiber diameter (d ~ 0.1 μm) (10), velocity at the mesh (U = 1.6 ± 0.6 cm·s⁻¹; mean ± SD), and seawater viscosity (ν = 0.83 × 10⁻⁶ m²·s⁻¹). Classic principles of low-Re filtration theory (9, 11) show that low-Re filter feeders can collect particles smaller than the mesh spacing by relying on mechanisms other than simple sieving. The primary mechanisms are direct interception of particles traveling on streamlines that come within one particle radius of the filter element, and diffusional deposition caused by Brownian effects or random motility, which deflect particles from streamlines and cause contact with the filter. Theoretical models of caddisfly larvae (12, 13) and experiments on marine appendicularians (14–16) showed encounter of particles much smaller than the mesh size via diffusional deposition and direct interception, and theory suggests that other encounter mechanisms (inertial impaction and gravitational deposition) are negligible for most marine filter-feeders (13, 17, 18). The transition from encounter to capture depends on the sticking coefficient α, which represents the fraction of encountered particles that is captured.

Empirical studies of salp retention efficiency found a size retention cutoff of 1–2 μm, but this remains inconclusive because submicrometer particles were neglected (4, 19) or undetectable (20). In fact, small cyanobacteria (0.7–1 μm) have been removed by salps during feeding studies (20) and identified in salp fecal pellets (3, 4). Because the smallest particles are the most abundant in the ocean (Fig. 1B) (29, 30), determining the encounter efficiency of submicrometer particles is of particular importance to quantify clearance rates and vertical transport of particulates. Contrary to the current understanding that salps do not retain particles below 1–2 μm, we show that salps can capture submicrometer particles, and do so at rates that exceed those of larger particles. We calculate that salps can fulfill their energetic requirements with particles smaller than the mesh width and propose that they can substantially influence particle-size spectra in the upper ocean, increasing particle size and thus accelerating vertical transport of particulate matter.

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Results

Epiﬂuorescence images revealed a regularly spaced rectangular feeding mesh (Fig. 2A) with a mean mesh width and length of $W = 1.5 \pm 0.5 \mu m$ and $L = 6.0 \pm 1.5 \mu m$ ($n = 9$; mean $\pm$ SD), respectively. Some strands were oriented obliquely or possibly tangled, but their number was small. Mesh width increased linearly with salp body length, $L_b$ (Fig. 2B), as expected from an isometric scaling. The use of a rectangular rather than a square mesh is common among aquatic ﬁlter feeders, including appendicularians and caddis ﬂy larvae, possibly optimizing the tradeoff between increasing encounter and lowering the mesh material and pressure drop (31).

Flow visualization provided both quantitative ﬂuid speeds near the ﬁlter and a qualitative picture of the feeding current. The mean speed ($U$) and maximum speed near the oral siphon were 1.6 and 3.8 cm·s$^{-1}$, respectively (Table 1). The mean speed was slightly lower than speeds measured just aft of the atrial (excurrent) siphon using particle-image velocimetry ($2.0–2.6$ cm·s$^{-1}$) (32), likely because the oral siphon has a larger cross-sectional area. Particle trajectories showed that opening of the oral siphon resulted in the intake of ﬂuid from around the edges of the siphon (Movie S1). Upon entering the pharyngeal chamber, water accelerated and then moved in a circular pattern, suggesting a tangential component of encounter between particles and the ﬁlter.

The observed feeding current speeds are much higher than those of appendicularians ($0.06–0.32$ cm·s$^{-1}$) (17, 33), which pump ﬂuid via sinusoidal motion of the tail, and doliolids ($0.11$ cm·s$^{-1}$) (34), which rely on cilia rather than muscles to draw ﬂuid toward a mucous ﬁlter; and are of the same order as feeding currents of copepods and krill ($0.6–1.5$ cm·s$^{-1}$) (35, 36), which generate ﬂow by the coordinated movement of feeding appendages. However, salps process much higher ﬂuid volumes than crustaceans, due to the considerably larger cross-sections of their feeding currents.

Table 1. Flow speed at *P. confoederata* feeding ﬁlter

<table>
<thead>
<tr>
<th>Individual</th>
<th>Stage</th>
<th>Body length, $L_b$ mm</th>
<th>Mean speed, $U$, cm·s$^{-1}$ ($n$)</th>
<th>Max speed, cm·s$^{-1}$</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Aggregate</td>
<td>27</td>
<td>$2.3 \pm 1.1$ (3)</td>
<td>4.1</td>
</tr>
<tr>
<td>2</td>
<td>Solitary</td>
<td>30</td>
<td>$1.2 \pm 0.9$ (9)</td>
<td>4.1</td>
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<tr>
<td>3</td>
<td>Solitary</td>
<td>34</td>
<td>$1.5 \pm 0.1$ (15)</td>
<td>2.4</td>
</tr>
<tr>
<td>4</td>
<td>Solitary</td>
<td>53</td>
<td>$1.9 \pm 1.0$ (13)</td>
<td>3.9</td>
</tr>
<tr>
<td>5</td>
<td>Solitary</td>
<td>56</td>
<td>$2.0 \pm 1.8$ (11)</td>
<td>6.7</td>
</tr>
<tr>
<td>6</td>
<td>Solitary</td>
<td>62</td>
<td>$0.8 \pm 0.2$ (14)</td>
<td>1.6</td>
</tr>
<tr>
<td>Mean ± SD ($n$)</td>
<td></td>
<td></td>
<td>$1.6 \pm 0.6$ (6)</td>
<td>$3.8 \pm 1.7$ (6)</td>
</tr>
</tbody>
</table>

Values expressed as mean $\pm$ SD. The mean speed weighted by the number of measurements for each organism was $1.5 \pm 1.1$ cm·s$^{-1}$. $n$, no. of measurements.

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**Fig. 1.** Pelagic tunicates and particulate food. (A) Schematic of three *Pegea confoederata* individuals (aggregate stage). Mucous feeding ﬁlter (normally transparent) is shaded in red, and direction of feeding current shown with arrows. (B) Size distribution of living and nonliving particles in the upper ocean, including viruses (21), colloids (22), submicron particles (23), bacteria (24, 25), Prochlorococcus (26), Synechococccus (25), nanoplanckton (24, 27), and microplankton (24, 27). Line is regression of microphytoplankton concentration vs. cell diameter, $\log_{10} C = -0.91 \log_{10}(d_P^3 \pi/6) + 3.5$; $C$ (particles·mL$^{-1}$), $d_P$ (μm) (28). Graphic by E. P. Oberlander, Woods Hole Oceanographic Institution.

**Fig. 2.** Filtering mesh of *P. confoederata*. (A) Epiﬂuorescent image of mesh. (Scale bar: 5 μm.) (B) Mesh width, $W$ (μm), as a function of body length, $L_b$ (mm; $n = 9$). The line corresponds to $W = 0.02L_b + 0.58$ ($n = 9$; $r^2 = 0.70$).
For example, grazing pressure by a bloom of the salp *Salpa thompsoni* in the Southern Ocean was equivalent to more than 100% of daily primary production, whereas grazing by dominant copepod species was negligible (37).

Both diffusional deposition and direct interception play a role in determining particle encounter by the filtering mesh, but direct interception is dominant for particle sizes $d_p > 0.05$ μm (Fig. 3). For $d_p = 0.01-0.05$ μm (viruses, colloids), diffusion is the primary mechanism of particle encounter, although efficiency is <2%. For the smallest particles, Brownian motion results in higher encounters compared with motility, whereas for $d_p > 0.2$ μm, diffusional deposition is larger for motile microorganisms. However, swimming is unlikely for organisms smaller than 0.6 μm, as Brownian rotation would turn them too frequently for swimming to be effective (39). For $d_p > 0.05$ μm, particles are more efficiently encountered via direct interception: for 0.5-μm nonmotile particles, encounter by direct interception is 254-fold higher than by diffusional deposition, and for 1-μm motile particles, that increase is 41-fold.

Because there are substantially higher numbers of small particles in the ocean (Fig. 1B), these particles can be disproportionately ingested even when encounter efficiencies are relatively low. Estimates of particle encounter based on encounter efficiency (Fig. 3) and realistic particle concentrations (Fig. 1B) show that, on average, particles in the 0.01- to 0.1-μm size range (viruses, colloids) are encountered at ~200x the rate of particles in the 0.1- to 1-μm range (submicron particles, bacteria, Prochlorococcus; Fig. 4A). However, larger particles still contribute more volume and carbon (Fig. 4B). The mean carbon contribution from 0.1- to 1-μm particles is 38x larger than from 0.01- to 0.1-μm particles. However, 1- to 10-μm particles contribute just 4x as much carbon as 0.1- to 1-μm particles (Fig. 4B). If only the outer 0.1 μm of each particle is digested, the situation is reversed: the 0.1- to 1-μm size range contributes 20% more carbon than the 1- to 10-μm range, and the maximum carbon contribution comes from 1.1-μm particles (Fig. 4B).

The model shows that particles smaller than the mesh width, $W = 1.4$ μm, supply a total of 0.15 mg C·h⁻¹ to a salp. The carbon ingestion rate of a 40-mm-long *P. confoederata* is 2.2% of the body carbon content each hour (41), or 0.02 mg C·h⁻¹ based on the carbon-to-body-length relationship of Madin et al. (42).

Therefore, even assuming that the sticking coefficient is small $(\alpha = 0.1-0.2)$, the carbon supplied by particles smaller than the mesh opening can support the majority or entirety of the organism’s carbon requirement.

To support this conclusion, predicted encounter rates via direct interception were tested experimentally by offering particles of three sizes ($d_p = 0.5$, 1, and 3 μm) to freshly collected *P. confoederata* and quantifying the relative capture rate of particles of each size. The particle size range where diffusional deposition is predicted to contribute significantly to encounter rates ($d_p < 0.05$; Fig. 3) was not tested in experiments, but its contribution in terms of carbon supply was predicted to be negligible based on model results (Fig. 4). When the same concentration of each particle size was offered, capture rates were similar among sizes, with a slight preference for the larger particles (Fig. 5A). Relative capture rates were 29.1 ± 8.6%, 30.1 ± 5.4%, and 40.8 ± 12.9% (mean ± SD) for 0.5-, 1-, and 3-μm particles, respectively. They were in general agreement with relative encounter rates from direct interception (relevant for particles $>0.05$ μm; Fig. 3), predicted to be 13.8%, 32.9%, and 53.3%, respectively. The discrepancy at the smallest size suggests that the contribution of smaller particles is even more pronounced than the model predicts. A model of simple sieving (17, 43) was an inferior predictor of relative encounter rates and was particularly poor at predicting encounter rates of the smallest particles, with mean relative encounter rates of 3.7%, 14.5%, and 81.9% for $d_p = 0.5$, 1, and 3 μm, respectively.

Offering a suspension of particles skewed toward higher concentrations at the smallest sizes confirmed these findings: measured rates were similar to those predicted by the direct interception model, and very different from the simple sieving model (Fig. 5B). In this case also, experiments showed an even higher capture rate of smaller particles than anticipated from modeled encounter rates. This difference could be due to a size dependence of the sticking coefficient $\alpha$, for example, due to larger drag forces experienced by larger particles (44).

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**Fig. 3.** Particle encounter efficiency predicted for *P. confoederata* over a range of particle sizes. Efficiency of direct interception (blue) is shown for the mean measured mesh width $W = 1.4$ μm (solid line), with lower and upper bounds (dashed lines) corresponding to minimum and maximum mesh widths ($W = 0.5$ and 2.3 μm, respectively; Fig. 2B). Efficiency of diffusional deposition is shown in green for passive particles and, whereas for $d_p > 0.2$ μm, diffusional deposition is larger for motile microorganisms. However, swimming is unlikely for organisms smaller than 0.6 μm, as Brownian rotation would turn them too frequently for swimming to be effective (39). For $d_p > 0.05$ μm, particles are more efficiently encountered via direct interception: for 0.5-μm nonmotile particles, encounter by direct interception is 254-fold higher than by diffusional deposition, and for 1-μm motile particles, that increase is 41-fold.

**Fig. 4.** Combined encounter rate predicted for direct interception and diffusional deposition (passive and motile particles) as a function of particle diameter for *P. confoederata*. Calculation based on Eq. 1, with $E$ from Fig. 3; $Q = 1.69$ mL·s⁻¹ and $\log_{10}C_c = -0.91 \log_{10}(d_p/\alpha) + 3.5$ (28) (Fig. 1). (A) Particle encounter rate and (B) carbon encounter rate based on $C_c = 0.11$·mg·cm⁻³, where $C_c$ is carbon content (mg·cell⁻¹), and $V$ is particle volume (μm³) (40). For the latter, two cases were considered: that the full particle is digested (solid line) or that only the outer 0.1-μm-thick shell of each particle is digested (dashed line). Note that above $d_p = 1.2$ μm, direct interception efficiency is 100%.
loidal components can be rich in polysaccharides, proteins, and lipids (46, 47), and can play an important role in biogeochemical processes (48–50). Although a conclusive understanding of the bioavailability of colloidal particles remains a major frontier for biogeochemists, work conducted in several aquatic ecosystems has shown that colloids are 6–37% organic carbon (median = 27%) (47). This finding is consistent with, and even somewhat on the larger side of, the figure for carbon content used here [−11%, based on C_{org} = 0.1110.80 (40) and a colloid density of 1.0-1.11 g·mL^{−1} (51)]. Regardless of the nutritional value, salps influence the turnover of the colloidal fraction of DOC through encounter and, ultimately, assimilation or defecation.

Salp filtration rates are among the highest in the ocean, reaching up to 15.3 mL·s^{−1} (2, 52), yet pelagic tunicates have among the smallest diameter mesh elements (see figure 6 in ref. 53) and mesh spacing (10) of all marine filter feeders. By constantly pumping large volumes of seawater through their bodies and retaining micrometer scale and submicrometer particles, salps are well adapted for existence in the oligotrophic ocean. Most salp species are more oceanic than neritic in distribution, and high particle concentrations in coastal areas can clog their filtering apparatus and disrupt feeding (54). Oceanic waters are frequently dominated by plankton that is too small to be captured by sieving. The finding that salps can fulfill their energetic requirements with only submicrometer particles helps explain this geographic distribution.

Carbon in the euphotic zone is typically regenerated on the order of hours via the microbial loop (55). Salps and other pelagic tunicates remove particles that are four to five orders of magnitude smaller than themselves, thereby bypassing several trophic levels (55). In addition, muscular pumping achieves a high throughput of seawater and associated particles compared with the much slower feeding currents generated by flagella or cilia in other planktonic filter feeders. Particles are packaged into membrane-bound fecal pellets that are often incompletely digested and therefore rich in carbon, nitrogen, and phosphorous (56), and contain trace elements (e.g., Ca and Mg) (4). Fecal pellets sink quickly and are transferred to a longer-lived pool in deeper water, where material is sequestered on time scales of years to centuries. The efficiency with which salps repackaged and export carbon from surface waters suggests that salps, particularly in bloom proportions, can profoundly influence biogeochemical cycling, as indicated also by a recent proposition to increase global salp populations to mitigate climate change (57). In summary, the high filtration rates of small particles imply that salps can rapidly transfer carbon and energy from the submicron size range of the particle spectrum to higher trophic levels by grazing, and to larger depths via their rapidly sinking fecal pellets. As such, salps can provide a substantial shortcut to flocculation in determining the contribution of small particles to vertical transport of particulate matter.

Materials and Methods

Specimen Collection. P. confoederata were collected in individual 800-mL plastic jars using blue-water SCUBA techniques (58) at the Liquid Jungle Lab off the Pacific coast of Panama (7° 50’ N, 81° 35’ W) during January 2007, 2008, and 2009. Animals were maintained in collection jars or in tanks (6–11 L) of field-collected seawater at in situ temperatures (26–28°C). All measurements were made within 12 h of collection.

Measurements of Mesh Size and Flow Speed. Filter mesh measurements were obtained by epifluorescence microscopy. Part of the mesh of P. confoederata was removed by gently inserting an ∼1 × 1-mm section of a glass coverslip through the oral siphon and sweeping it through the pharyngeal chamber using forceps. After adding 50–100 μL of lectin-fluorescein isothiocyanate in seawater solution (1 mg·mL^{−1}), the mesh was imaged using a Zeiss Axiosstar Plus microscope with an HBO 50 epifluorescence lamp, a 100× objective, and a Nikon Coolpix 8800 camera. This is the first time the filtering mesh was imaged using a wet preparation to reduce sample distortion caused by drying and shrinking associated with TEM and SEM techniques (3, 59). Data were
acquired from six P. confederata solitary and three aggregates, ranging from 16 to 60 mm long. Mesh length, L, and width, W, were measured in ImageJ (http://rsbweb.nih.gov) for multiple mesh openings (mean ± SD was 16 ± 10) and averaged for each individual.

The flow pattern and speed were determined using particle tracking. Individual P. confederata were placed in custom-built acrylic tanks with field-collected seawater seeded with 10 ± 2 μm titanium dioxide particles. Particles were illuminated with a 1-mm-thick laser sheet (30 mW, 500 nm wavelength) generated using a Powel lens (Lasiris) and their motion videotaped with a Sony HDR-HC7 videocamera (1,440 × 1,080 pixels, 30 fps). Because salps are transparent, particles could be tracked within the pharyngeal chamber until contact with the filtering mesh occurred. Velocities were determined by tracking individual particles between frames relative to landmarks on the salp body or by measuring particle streak lengths in a single frame using ImageJ.

Particle Encounter Model. The encounter rate (60)

\[ P = B \cdot C = E \cdot Q \]  

is the product of the encounter rate kernel, \( B \) (mL⁻¹s⁻¹), and the particle concentration, \( C \) (particles mL⁻¹). Here, \( B = E \cdot Q \), where \( E \) (dimensionless) is the capture efficiency (Si Appendix) and \( Q \) (mL⁻¹s⁻¹) is the volume flow rate through the salp. Both \( E \) and \( Q \) depend on particle diameter, \( d_p \). Particle capture by salps is a low Re-number process, indicating that viscous forces dominate inertial forces in determining capture. The flow through the mesh has \( Re = \frac{d_P \cdot \nu \cdot \rho}{\mu} = 3 \times 10^2 \), and the flow around an individual mesh strand (diameter \( d \sim 0.1 \mu m \)) has \( Re = \frac{d \cdot \nu \cdot \rho}{\mu} = 2 \times 10^3 \). Particle inertia is negligible, as the Stokes number \( d_P^2 \cdot \rho \cdot u / 18 \pi \eta \) is always less than 1 for \( d_P < 10 \mu m \), particle density \( \rho = 1,037 \) kg m⁻³, and seawater density \( \rho = 1,030 \) kg m⁻³. Thus, particle capture is limited to noninertial mechanisms, which include direct interception and diffusional deposition (12).

We used a model for capture efficiency, \( E \), by a rectangular mesh (Si Appendix) (12), with parameters that were directly measured (mesh dimensions, flow through the filter) or taken from literature (mesh fiber diameter, particle size distribution). We assumed spherical particles in Eq. 1. The encounter of nonmotile and motile particles by diffusional deposition was modeled by a diffusivity based on Brownian motion and random motility, respectively (Si Appendix).

The volume flow rate through the salp, \( Q = 1.69 \) mL s⁻¹, was determined as the average from three studies (20, 52, 61) and had an SD of 1.44 mL s⁻¹. The particle size distribution, concentration \( C \) of particles of size \( d_P \), was obtained from four Atlantic Ocean transects (28) and is likely a conservative estimate, as other studies found higher concentrations in all size ranges (Fig. 1B). Carbon capture was calculated using the relation \( C_{\text{ECO}} = 0.114 d^{-0.69} \) between carbon content, \( C_{\text{ECO}} \) (μg cell⁻¹), and particle volume, \( V \) (μm³), for phytoplankton (similar relations apply for bacterioplankton and colloid) (47, 62). Because partially digested undigestible particles are frequently observed in salp fecal pellets (3, 4), we also explored the implications for carbon encounter if only the outer 0.1 μm of each particle is digested. Relative estimates of particle and carbon encounters mentioned in the text were computed based on uniformly distributed values of particle diameter with spacing of 0.01 μm.

Particle Capture Experiments. Relative retention efficiencies of \( d_P = 0.5, 1 \), and 3 μm fluorescent polystyrene microspheres (Polysciences, Inc.) were determined using two feeding experiments, performed within 3 h of specimen collection. Microspheres were pretreated with 5 mg mL⁻¹ BSA for 12–48 h to avoid clumping (63). In the first experiment, microspheres were added to each jar at a concentration \( C = 10^5 \) μm⁻¹ for each size. After 2 h, P. confederata guts were excised and ground using a mortar and pestle along with several microliters of seawater. Two 2-μL subsamples of the homogenate were examined using epifluorescence microscopy at 200x magnification and 365 ± 12 nm excitation, and particles of each size were counted from three fields of view from each 2-μL subsample. The three particle sizes were distinguished based on size (\( d_P = 0.5, 1, \) and 3 μm), and emission wavelength (486, 407, and 486 nm, respectively). Each count included a minimum of 50 particles. In the second experiment the starting concentrations were \( C = 10^3, 10^4, \) and \( 10^5 \) μm⁻¹ for \( d_P = 0.5, 1, \) and 3 μm, respectively, to better represent the presence of small particles in the ocean (Fig. 1B). For both experiments, relative retention efficiencies were determined by dividing the count for a given particle size by the total count for all three sizes. Comparisons were made between relative retention efficiencies from experiments, the low-Re encounter model, and a simple sieving model based on an experimentally determined Gaussian distribution of mesh widths (17, 43).

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