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**Title:** Protocol for Sampling and Analysis of Finished Medical Marijuana Products and Marijuana-Infused Products for Massachusetts Registered Medical Marijuana Dispensaries

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The Protocol contains the following sections:

- 1.0 Purpose and Applicability
- 2.0 Definitions and Acronyms
- 3.0 Applicable Regulations
- 4.0 Sampling and Analysis Requirements
- 5.0 Sampling Program Design
- 6.0 Sample Collection Procedures
- 7.0 Sample Analysis
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## **1.0 Purpose and Applicability**

### **1.1 Purpose**

The purpose of this Protocol is to provide Massachusetts Registered Marijuana Dispensaries (RMDs) with required and recommended best practices for the collection and analysis of plant material and other finished medical marijuana products and marijuana-infused products (MIPs) to comply with Massachusetts regulation 105 CMR 725.000, Implementation of an Act for the Humanitarian Medical Use of Marijuana.

This protocol is subject to revision based on evolving best practices, updated scientific information or standards/guidelines, or other information relevant to the contents of the protocol.

### **1.2 Applicability**

This protocol applies only to Massachusetts RMD operations, and not hardship cultivation operations. Testing requirements in the protocol apply only to the medical marijuana products dispensed by Massachusetts RMDs, including finished medical marijuana (i.e., plant material, resin, concentrates) and MIPs made with finished medical

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marijuana ingredients. The protocol only addresses sampling and analysis to characterize cannabinoid identity and content profiles, and biological (microbial and fungal) and chemical (e.g., solvents, pesticides, growth enhancers, metals) contaminants introduced through cultivation of marijuana plants and post-harvest processing and handling of marijuana products and ingredients.

This protocol does not apply to nutritional product testing, allergen testing, or characterization of non-marijuana ingredients in MIPs. It does not address sampling and analysis to verify compliance with state regulations or best practices for production and handling of food products, pharmaceuticals, or dietary supplements, except for criteria for biological and chemical contaminants that may be introduced through inclusion of medical marijuana as an ingredient.

Sampling and analysis of environmental media used for cultivation are addressed in a companion protocol, *Protocol for Sampling and Analysis of Environmental Media for Massachusetts Registered Medical Marijuana Dispensaries*.

## 2.0 Definitions and Acronyms

Terms listed in italic typeface are those defined in 105 CMR 725.004. Additional terms defined for this protocol are not in italic typeface.

**Cannabinoid** means any of several compounds produced by marijuana plants that have medical and psychotropic effects.

**Cannabinoid Profile** means amounts, expressed as the dry-weight percentages, of  $\Delta^9$ -tetrahydrocannabinol ( $\Delta^9$ -THC) and cannabidiol (CBD) in a medical marijuana product. Amounts of other cannabinoids may be reported, but are not required.

**Cannabis Concentrate** means a marijuana product derived by using solvents to extract and concentrate cannabinoid compounds. Concentrates are typically in the form of oils, pastes, waxes, or solids.

**Cannabis Resin**, commonly known as “hashish,” “hash,” or “bubble hash,” means a solid medical marijuana product produced by gathering and compressing the cannabinoid-rich trichomes (i.e., keif) of the marijuana plant.

**Certificate of Registration** means the certificate issued by the Department that confirms that a RMD has met all requirements pursuant to the Act and 105 CMR 725.000 and is registered by the Department.

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**Cultivation Batch** means a collection of marijuana plants from the same seed or plant stock and that are cultivated and harvested together. Because they are cultivated in the same location and time, plants in a cultivation batch receive an identical propagation and cultivation treatment (e.g., growing media, ambient conditions, watering and light regimes, agricultural or hydroponic inputs). The RMD must assign and record a unique, sequential alphanumeric identifier to each cultivation batch for the purpose of production tracking, product labeling, and product recalls.

**Department or DPH** means the Massachusetts Department of Public Health.

**Dispensary Agent** means a board member (including advisory board members), director, employee, executive, manager, or volunteer of a RMD, who is at least 21 years of age. Employee includes a consultant or contractor who provides on-site services to a RMD related to the cultivation, harvesting, preparation, packaging, storage, testing, or dispensing of marijuana.

**Edible Marijuana-Infused Products (edible MIPs)** means a marijuana-infused product (MIP) that is to be consumed by eating or drinking.

**Duplicate Samples** means two samples taken from and representative of the same material that are carried through all steps of the sampling and analytical procedures in an identical manner. Duplicate samples may be used to evaluate variance in the evaluation method, including sampling and analysis.

**Finished Medical Marijuana** means usable marijuana, cannabis resin, or cannabis concentrate.

**Finished Plant Material** means usable marijuana that has been trimmed and dried. Trimming includes removing the leaves immediately subtending the buds as well as any dead leaves or stems.

**Flowering** means the gametophytic or reproductive state of marijuana in which the plant produces flowers, trichomes, and cannabinoids characteristic of marijuana.

**Hardship Cultivation Registration** means a registration issued to a registered qualifying patient under the requirements of 105 CMR 725.035.

**Marijuana** means all parts of the plant *Cannabis sativa* L., whether growing or not; the seeds thereof; and resin extracted from any part of the plant; and every compound, manufacture, salt, derivative, mixture, or preparation of the plant, its seeds or resin. It does not include the mature stalks of the plant, fiber produced from the stalks, oil, or cake made from the seeds of the plant, any other compound, manufacture, salt, derivative, mixture, or preparation of the mature stalks, except the resin extracted

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therefrom, fiber, oil, or cake or the sterilized seed of the plant which is incapable of germination. The term also includes MIPs except where the context clearly indicates otherwise.

**Marijuana-Infused Product (MIP)** means a product infused with marijuana that is intended for use or consumption, including but not limited to edible products, ointments, aerosols, oils, and tinctures. These products, when created or sold by a RMD, shall not be considered a food or a drug as defined in M.G.L. c. 94, s. 1.

**Registered Marijuana Dispensary (RMD)** means a not-for-profit entity registered under 105 CMR 725.100, that acquires, cultivates, possesses, processes (including development of related products such as edible MIPs, tinctures, aerosols, oils, or ointments), transfers, transports, sells, distributes, dispenses, or administers marijuana, products containing marijuana, related supplies, or educational materials to registered qualifying patients or their personal caregivers. Unless otherwise specified, RMD refers to the site(s) of dispensing, cultivation, and preparation of marijuana.

**Mycotoxin** means a secondary metabolite of a microfungus that is capable of causing death or illness in humans and other animals. For the purposes of this regulation mycotoxins include alfatoxin B1, alfatoxin B2, alfatoxin O1, alfatoxin O2, and Ochratoxin A.

**Production Batch** means a batch of finished plant material, cannabis resin, cannabis concentrate, or MIP made at the same time, using the same methods, equipment, and ingredients. The RMD must assign and record a unique, sequential alphanumeric identifier to each production batch for the purpose of production tracking, product labeling, and product recalls. All production batches must be traceable to one or more marijuana cultivation batch(es).

**Propagation** means the reproduction of marijuana plants by seeds, cuttings, or grafting.

**Residual Solvent** means a volatile organic chemical used in the manufacture of a medical marijuana product and that is not completely removed by practical manufacturing techniques.

**Tincture** means an extract, typically in ethanol, of usable marijuana. Marijuana tinctures sometimes are made with glycerin or other alternatives to ethanol.

**Trichome** means a cannabinoid-producing glandular structure that grows on the plant surface of marijuana plants, particularly on the buds of the female plant.

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**Usable Marijuana** means the fresh or dried leaves and flowers of the female marijuana plant and any mixture or preparation thereof, including MIPs, but does not include the seedlings, seeds, stalks, or roots of the plant.

**Vegetation** means the sporophytic state of the marijuana plant, which is a form of asexual reproduction in plants during which plants do not produce resin or flowers and are bulking up to a desired production size for flowering.

## 3.0 Applicable Regulations

This protocol was developed to provide RMDs with guidance on complying with the 105 CMR 725.000 regulations. In particular, the detailed steps outlined in this protocol address requirements of the following sections of the regulations. RMDs should be familiar with the applicable regulations to ensure full compliance.

- 725.105(A)(7) Requirement of plans for quality control, including product testing for contaminants
- 725.105(B) Cultivation, acquisition, and distribution requirements
- 725.105(B)(2) Marijuana obtained from another RMD
- 725.105(C) Requirements for handling and testing marijuana and for production of MIPs
- 725.105(E)(2) Labeling of marijuana
- 725.300(E) Testing pursuant to DPH inspection

## 4.0 Sampling and Analysis Requirements

Sampling and analysis requirements apply to all marijuana-containing products dispensed by Massachusetts RMDs, which may include finished plant material, cannabis resin, cannabis concentrates, and MIPs. Because the nature and concentrations of contaminants and cannabinoid compounds may change throughout the production process, from cultivation through packaging, this section identifies the types of sampling and analysis that are required for each type of product. The results of the sampling and analysis are required for both quality control and labeling requirements (e.g., cannabinoid profile, testing certification). RMDs must ensure and be able to demonstrate to inspectors, that product label information has been verified for all products.

#### **4.1 Overview of Medical Marijuana Products and their Production**

Medical marijuana products that may be dispensed by RMDs in Massachusetts include finished plant material, cannabis resin, cannabis concentrates, and a variety of MIPs. Except in the case of emergency situations, the marijuana for all of these product categories must originate with plants cultivated by the RMD operator (105 CMR 725.105(B)) and all product labeling must include a batch number to identify the batch associated with manufacturing and processing (105 CMR 725.105(E)). Therefore, RMDs are responsible for carefully tracking medical marijuana throughout the production cycle, from cultivation through dispensing to patients. Medical marijuana procured by a RMD from another RMD pursuant to 105 CMR 725.105(B)(2) must be tested by the supplying RMD and documentation of testing consistent with this protocol must be provided to the receiving RMD by the supplying RMD, along with chain-of-custody documentation.

Exhibit 1 provides an overview of the medical marijuana production process as regulated in Massachusetts. During cultivation, plants are typically grown from seed, cuttings, or through a tissue culture method called micropropagation (AHP 2013). Plants may be grown in soil, other solid growth media, or in hydroponic systems. All cultivation methods place the plants in contact with environmental media and other inputs, such as soil or agricultural products, which have the potential to introduce chemical or biological contaminants.<sup>1</sup> Because medically-active compounds are at their highest concentration on the inflorescences of the female plant, marijuana plants are harvested when the plants reach peak maturity. Post-harvest handling steps include drying and trimming, which should be managed carefully to avoid mold and bacterial growth and to preserve medicinally-active compounds. For further details on medical marijuana cultivation and post-harvest handling methods, refer to AHP (2013).

Harvested and dried marijuana plants can be used directly to produce any of the three finished medical marijuana types. Dried and trimmed usable marijuana, most importantly the inflorescences (i.e., “buds”), may be used directly (e.g., smoked) as a medical product without further processing. It also may be used as a source material for other finished marijuana products or as an ingredient in MIPs. Cannabis resin, commonly referred to as “hashish” or “hash,” is formed by collecting and compressing cannabinoid-containing resin glands (i.e., trichomes). Cannabis resin also includes “bubble hash,” which is made by extracting the resin glands using cold water and physical separation (Colorado Pot Guide, 2014). Concentrates, which include various oils, waxes, and solids, are produced with solvent extraction methods. Concentrates have higher

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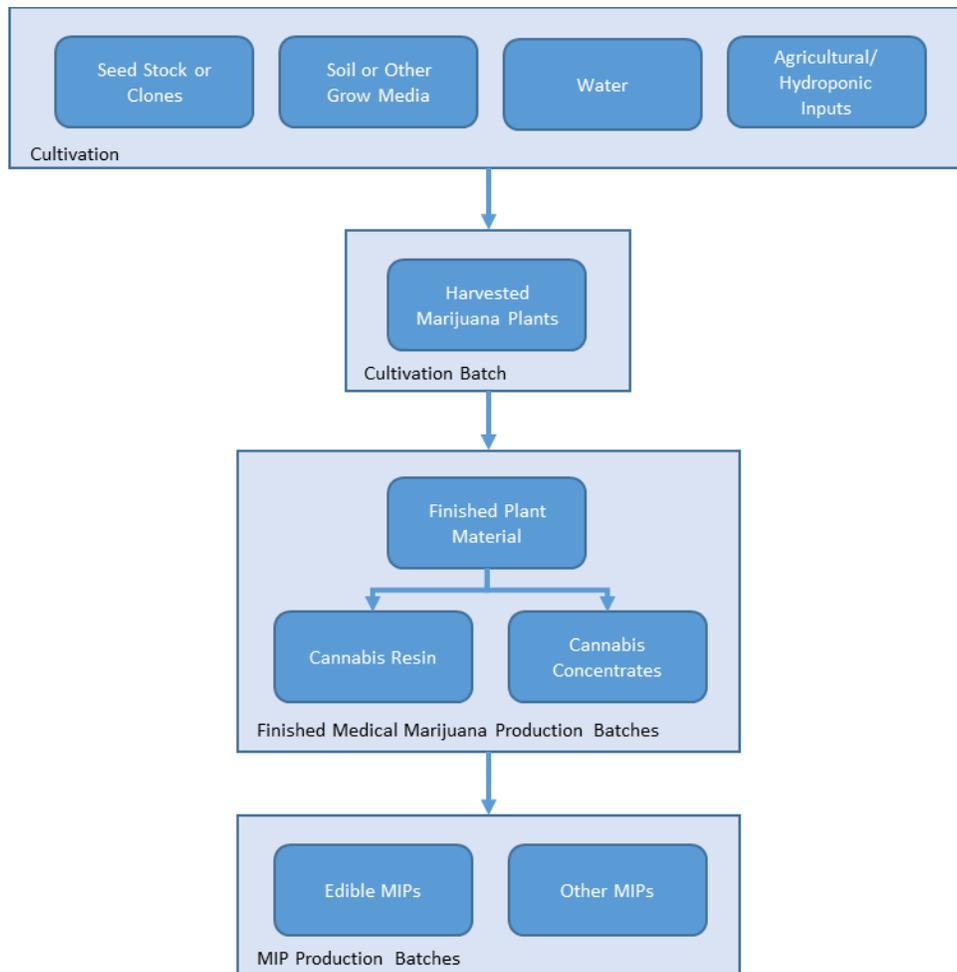
<sup>1</sup> Testing for media used in marijuana cultivation is discussed in the companion *Protocol for Sampling and Analysis of Environmental Media for Massachusetts Registered Medical Marijuana Dispensaries*.

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cannabinoid concentrations than other finished marijuana products, but also may contain residuals of potentially harmful solvents if not manufactured properly. In addition, any contaminants present in the source plant material may be concentrated in a resin or concentrate product.

## Exhibit 1. Overview of Medical Marijuana Production



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Under 105 CMR 725.004, a MIP is defined as “a product infused with marijuana that is intended for use or consumption, including but not limited to edible products, ointments, aerosols, oils, and tinctures.” MIPs available to patients and consumers may include, but are not limited to baked goods; lozenges and candies; teas and other beverages; creams and salves; tinctures; and products for vaporization.

## 4.2 Medical Marijuana Testing Requirements

Testing for finished medical marijuana and MIPs includes screening for chemical and biological contaminants (Section 4.2.1) and cannabinoid profile testing (Section 4.2.2). Section 4.2.3 discusses methods for determining the amount of usable marijuana contained within a dispensed product, as required for product labeling. Sections 5.0 through 7.0 further describe the detailed sampling frequency, sample collection procedures, and analyses required for contaminant and cannabinoid profile testing.

This protocol defines the minimum testing required to conform with the 105 CMR 725 regulations. RMDs have discretion to perform analysis beyond these requirements.

### 4.2.1 Contaminant Testing

Contaminant testing requirements are based on the contaminants potentially introduced at each stage of production. Exhibit 2 identifies the potential contaminants of concern during each stage of medical marijuana production and the testing requirements for each product type.

#### ***Cultivation***

Cultivation is not in the scope of testing of this protocol, but is included in Exhibit 2 to identify the contaminants of concern potentially introduced during cultivation. These include non-organic pesticides, metals, and other synthetic organic compounds in environmental media or other cultivation inputs (e.g., soil amendments, hydroponic products), as well as fungal and bacterial growth on the plants. Environmental media must be tested, as described in the *Protocol for Sampling and Analysis of Environmental Media for Massachusetts Registered Medical Marijuana Dispensaries*, to reduce the introduction of chemical contaminants during cultivation. However, this testing will not necessarily ensure that the marijuana plants are free of chemical contaminants, and does not address fungal/bacterial infestation. Therefore, medical marijuana products must be tested for chemical contamination before they can be distributed.

Marijuana should be cultivated and harvested in traceable “cultivation batches,” such that all marijuana within a cultivation batch has been produced with the same seed or plant stock, soil or other solid growing media, water, other agricultural/hydroponic inputs, and growing conditions. Cultivation batches should be sequentially numbered and traced throughout post-harvest production steps, and manufacturing/processing batch numbers

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must be included on the labels of all products to facilitate product recalls (105 CMR 725.105(E)(2)(e) and 725.105(E)(3)(g)).

### ***Finished Plant Material***

Finished plant material dispensed to patients consists of usable marijuana that has been trimmed and dried. Trimming includes removing the leaves immediately subtending the buds as well as any dead leaves or stems (AHP 2013). A “production batch” of finished plant material must be traceable to one or more cultivation batch(es). All production batches of finished plant material must be tested for pesticides and metals, which may be introduced during cultivation. Production batches intended for dispensing and direct use as a medical product must also be tested for biological contaminants (bacteria, fungi, and mycotoxins), as shown in Exhibit 2. Finished plant material is tested instead of living or freshly harvested plants because drying and trimming may affect the concentrations of contaminants and because fungal/bacterial growth may occur during finishing.

Finished plant material that exceeds a limit (see Section 7.0) for any contaminant included in the required testing cannot be distributed as finished medical marijuana.

### ***Cannabis Resins and Concentrates***

Cannabis resins and concentrates may be produced from the finished plant material of one or more cultivation batches. If the finished plant material fails to meet a required testing requirement, but the finished plant material is not dispensed to a patient, then it may be used to derive resins and concentrates. The resins and concentrates may be dispensed as long as they meet the respective concentration limit identified in Section 7.0. Each production batch of cannabis resin or concentrate must be given a sequential identifier for product tracking and labeling. The RMD must keep records of the marijuana cultivation batch(es) used for each production batch, and include the manufacturing/processing batch number on product labels.

Testing requirements for cannabis resins and concentrates are summarized in Exhibit 2. Because these products may be made only from plant material that has already tested below limits for pesticides, testing for these contaminants is not required again. However, cannabis concentrates must be tested for metals, as well as residual solvents if solvents were used in their production. Specifically, testing is required for any solvent used to make a cannabis concentrate production batch.

All cannabis resin or concentrate production batches intended for distribution to patients as finished medical marijuana products must be tested for bacteria, fungi, and mycotoxins. Testing for these biological contaminants is not required for cannabis resin or concentrate production batches that will be used only to manufacture MIPs.

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If required testing finds that a production batch of cannabis resin or concentrate exceeds any applicable contaminant limit (see Section 7.0), the production batch cannot be dispensed as a finished medical marijuana product.

### ***MIPs***

DPH assumes that all MIP production batches will be destined for dispensation and patient use. Therefore, all MIP production batches must be tested for biological contaminants (bacteria, fungi, and mycotoxins). Production batches must be discarded and not dispensed to patients if any biological contaminant limit is exceeded.

MIPs may be made only with finished medical marijuana products that have passed applicable metals, pesticide, and solvent testing requirements. For this reason, testing MIPs for metals, pesticide, and solvent contaminants is not required. However, RMDs have discretion to perform this testing of MIPs voluntarily.

Each MIP production batch must be given a sequential identifier (ID) for product tracking and labeling. Records must be kept that identify the cultivation batch(es) and finished medical marijuana production batches associated with each MIP production batch. The manufacturing/processing batch number must be included on product labels to aid in product tracking and recalls.

### **4.2.2 Cannabinoid Profile Testing**

All medical marijuana products, shown in Exhibit 1, including any finished medical marijuana or MIP, must bear a label that identifies the list of ingredients, including the cannabinoid profile of the marijuana contained within the product, including the THC level (105 CMR 725.105(E)(2)(f) and 725.105(E)(3)(e)). Therefore, for the purposes of labeling medical marijuana products in Massachusetts, the cannabinoid profile must include, at a minimum, the percentage by dry weight (i.e., the weight of the material remaining after it has been thoroughly dried) of  $\Delta^9$ -tetrahydrocannabinol ( $\Delta^9$ -THC) and cannabidiol (CBD). Medicinal benefits have been attributed to other cannabinoids, and these compounds may be included in the cannabinoid profile at the discretion of the RMD.

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**Exhibit 2. Contaminants of Concern and Testing Requirements for Medical Marijuana Products**

Production Stage/Product	Contaminants of Concern	Required Testing
Cultivation [see <i>Protocol for Sampling and Analysis of Environmental Media for Massachusetts Registered Medical Marijuana Dispensaries</i> ]	<ul style="list-style-type: none"> <li>• Non-organic pesticide residues, metals, and other organic chemicals (e.g., PCBs) taken up by plants from soil/growing media</li> <li>• Non-organic pesticides, metals, and other organic chemicals taken up by plants from water source</li> <li>• Non-organic pesticides applied to plants in noncompliance</li> <li>• Unidentified non-organic pesticides, metals, and other chemicals in non-pesticide agricultural or hydroponic products</li> <li>• Fungi/microbes growing on plants or equipment</li> </ul>	<ul style="list-style-type: none"> <li>• Test soil or other solid media for pesticides, metals, and other organic chemicals (e.g., PCBs)</li> <li>• Test water sources for pesticides, metals, and bacteriological contaminants</li> <li>• No fungi/mycotoxin testing</li> </ul>
Finished Plant Material	<ul style="list-style-type: none"> <li>• Contaminants remaining from cultivation</li> <li>• Fungal/microbial growth during finishing</li> </ul>	All Production Batches: <ul style="list-style-type: none"> <li>• Pesticides and plant growth regulators</li> <li>• Metals</li> </ul> Only Production Batches to be Dispensed as Medical Marijuana Product: <ul style="list-style-type: none"> <li>• Bacteria/fungi/mycotoxins</li> <li>• Cannabinoid profile</li> </ul>
Cannabis Resin	<ul style="list-style-type: none"> <li>• Contaminants remaining from cultivation</li> <li>• Fungal/microbial growth during production</li> </ul>	All Production Batches: <ul style="list-style-type: none"> <li>• Metals</li> </ul> Only Production Batches to be Dispensed as Medical Marijuana Product: <ul style="list-style-type: none"> <li>• Bacteria/fungi/mycotoxins</li> <li>• Cannabinoid profile</li> </ul>

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Production Stage/Product	Contaminants of Concern	Required Testing
Cannabis Concentrates	<ul style="list-style-type: none"> <li>• Contaminants remaining from cultivation</li> <li>• Solvents used to extract oil</li> <li>• Fungal/microbial growth during production</li> </ul>	<p>All Production Batches:</p> <ul style="list-style-type: none"> <li>• Solvents</li> <li>• Metals</li> </ul> <p>Only Production Batches to be Dispensed as Medical Marijuana Product:</p> <ul style="list-style-type: none"> <li>• Bacteria/fungi/mycotoxins</li> <li>• Cannabinoid profile</li> </ul>
MIPs	<ul style="list-style-type: none"> <li>• Contaminants in finished marijuana ingredients</li> <li>• Fungal/microbial growth during production</li> </ul>	<p>All Production Batches:</p> <ul style="list-style-type: none"> <li>• Bacteria/fungi/mycotoxins</li> <li>• Cannabinoid profile</li> </ul>

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It is important to note that heat (including combustion) can cause chemical reactions that convert cannabinoids to more or less potent forms. For example, combustion (e.g., during smoking) causes non-psychoactive cannabinoid acids, abundant in the plant material, to be converted to psychoactive forms. However, medical users report health benefits from products that do not require high temperatures or combustion for production or use (AHP 2013). Because production of finished medical marijuana products and MIPs may affect cannabinoid chemistry, as well as the concentration or dilution of active ingredients, each product type must be tested to characterize the cannabinoid content and profile.

### **4.2.3 Usable Marijuana Content**

105 CMR 725.105(E)(2)(c) and 725.105(E)(3)(d) require labels of medical marijuana products to identify the quantity of usable marijuana contained within the product, as measured in ounces. For finished plant material and products containing finished plant material the quantity of usable marijuana is simply the weight in ounces of the plant material in the product. Massachusetts has determined that 10 ounces of finished plant material is the maximum 60-day supply allowed for medical marijuana patients. This is the largest amount of usable medical marijuana that may be dispensed by any RMD in Massachusetts.

When finished plant material is used to derive cannabis resin or concentrates, processing alters the physical form and quantity (i.e., weight and volume) of the usable marijuana. To enable the comparison of usable marijuana in the various product types, DPH developed assumptions that should be used to express the quantity of usable marijuana in cannabis resins or concentrates in terms of the equivalent ounces of plant material. Based on sources reviewed by DPH, it can be assumed that the yield of a cannabis resin or concentrate is 15 percent of the starting weight of plant material. When the weight of cannabis resin or concentrate in a dispensed product is known, the quantity of usable marijuana, expressed in equivalent plant material weight, should be calculated by multiplying the resin or concentrate weight by 6.7 (i.e.,  $1 \div 0.15$ ). For example, the quantity of usable marijuana in 1.5 ounces of cannabis oil is 10 ounces (1.5 ounces of cannabis oil  $\times$  6.7 = 10 ounces of usable marijuana). Therefore, 1.5 ounces of cannabis oil is equivalent to the maximum 60-day supply of useable plant material.

The amount of usable marijuana in a MIP is equal to the amount of usable marijuana included in the product ingredients, measured before mixing, baking, or other processing or manufacturing steps. If more than one type of finished marijuana ingredient is used to prepare a MIP, the amount of usable marijuana in the MIP is the sum of the usable marijuana in the ingredients.

## **5.0 Sampling Program Design**

Under 105 CMR 725.105(C), medical marijuana must be tested for the cannabinoid profile and contaminants. The medical marijuana products to be tested include: finished plant material (i.e., inflorescences or “buds”), cannabis resin, cannabis concentrates, and various types of MIPs. The purpose of testing is to ensure product quality and safety, and to provide information needed for product labeling requirements.

Because it is not possible to test all medical marijuana, RMDs must collect representative samples to provide to the analytical laboratory. Specifically, each medical marijuana production batch must be sampled and analyzed, and the samples collected for a production batch must be representative of all of the medical marijuana in the batch. The protocol provides the following definition of production batch:

***Production Batch** means a batch of finished plant material, cannabis resin, cannabis concentrate, or MIP made at the same time, using the same methods, equipment, and ingredients. The RMD must assign and record a unique, sequential alphanumeric identifier to each production batch for the purpose of production tracking, product labeling, and product recalls. All production batches must be traceable to one or more marijuana cultivation batch(es).*

Samples from each production batch must be collected in a ready-to-use condition. For production batches that will be dispensed to patients, ready-to-use means ready for packaging or post-packaging. For other production batches, ready-to use means ready for use as an intermediate or ingredient in making other products. After samples are collected, the entire production batch must be stored in a secure, cool, and dry location until analytical results are returned by the laboratory.

Sampling frequency is dictated by the production schedules, which may vary among RMDs due to scale, product types dispensed, and patient demand. The RMD is responsible for implementing a production batch tracking approach that meets the regulatory needs and definitions as well as ensuring representative sample collection and analysis of those batches. The RMDs must be able to demonstrate to inspectors that the production tracking, sampling, and analysis procedures are capable of obtaining representative samples. The guidelines below are provided to aid RMDs in developing an approach that meets DPH requirements for representativeness.

To perform required testing, RMDs will collect samples to be analyzed by independent and appropriately certified laboratories, as noted in Section 7 of this protocol. The amounts of sample required for cannabinoid or contaminant testing may vary by analytical method and laboratory-specific procedures, therefore the RMD should confer with the laboratory to determine the minimum sample size required for evaluation. In all

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cases, the amount of sample supplied to the laboratory should be large enough and sufficiently homogenized to provide a representative sample of the production batch but not in excess to raise issues with possible diversion or waste disposal.

### 5.1 Representative Sampling

Specific procedures for collecting representative samples of medical marijuana production batches are likely to vary depending on several attributes of the products and production methods:

**Homogeneity** – A sample is more likely to accurately represent the production batch if the material is homogenous (i.e., well mixed). Mixing or other homogenization steps help to homogenize the product before sample collection.

**Physical Form** – Production batches will vary in physical form (e.g., liquids, solids), density, and viscosity. Physical form can affect homogeneity, homogenization steps, and sample collection methods. For example, liquid products can be homogenized by stirring. Grinding and other methods described further below can be used to homogenize solid products.

**Quantity** – Because production batches may vary in scale (i.e., volume or weight), varying numbers or sizes of samples may be required to promote representativeness.

In addition, sample representativeness can be affected by the timing and frequency of sample collection. Because of variation among production schedules (e.g., due to product type, production scale, patient demand), sampling frequencies will vary among RMDs and production batches. However, representativeness will be ensured by the requirement that all production batches are tested.

### 5.2 Representative Sampling by Physical Form and Quantity

Exhibit 3 provides instructions for representative sampling of medical marijuana production batches, including finished medical marijuana products and MIPs. These instructions were developed based on sampling guidance for food products and herbal medicines developed by the Codex Alimentarius Commission (1999) and the United States Pharmacopeia Chapter 561 (USP, Undated-b), respectively, and account for differences in the physical forms of the production batches as they relate to homogeneity and quantity. If application of these guidelines is impractical for specific products, it is the responsibility of the RMD to develop and document a scientifically-defensible sampling approach.

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Homogeneity plays an important role in methods for representative sampling. While liquid products such as cannabis oil and liquid MIPs can be stirred or mixed to homogenize the product before sampling, other products such as cannabis resin, baked goods, or hard candies cannot. Homogenization of some solid products, such as ground plant material or semi-solid resin is possible. Because of its importance, further guidance on homogenization methods is provided in Section 5.3.

### Exhibit 3. Representative Sample Program Design

Homogeneity	Mass of Production Batch (kg)	Minimum Number of Samples/subsamples	Collection Method
Production batch can be homogenized or assumed to be well mixed or homogenous	Any production batch mass	1 sample	<ul style="list-style-type: none"> <li>• Mix product</li> <li>• Withdraw amount sufficient for evaluation</li> <li>• Submit to laboratory for analysis</li> </ul>
Production batch cannot be homogenized or is of unknown homogeneity	≤1	3 subsamples	<ul style="list-style-type: none"> <li>• Mix product to degree possible</li> <li>• Withdraw subsamples from different areas (such as lower, middle, and upper portions of container)</li> <li>• Composite subsamples, if possible, to amount sufficient for evaluation</li> <li>• Submit to laboratory for analysis</li> </ul>
	1-5	5 subsamples	
	≥5	10 subsamples	

Sources: USP Chapter <561> (Undated) and Codex Alimentarius Commission (1999)

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### 5.3 Sampling Guidance by Matrix

Finished marijuana products and MIPs can be in varied physical states or matrix (e.g., liquids to hard solids). To better understand the specific requirements the following guidance is provided based on the matrix of the material to be characterized.

#### ***Liquids (Cannabis Oil and Some MIPs)***

Liquid products such as cannabis oil or liquid MIPs should be thoroughly stirred or mixed before sampling to ensure homogenization of the sample. Cannabis oil or other liquid cannabis from each production batch should be sampled using units of volume. Samples of concentrates or oils should be collected following each production batch if they are to be sold, and before any further processing into MIPs.

#### ***Finished Plant Material or Friable MIPs***

Sampling shall be performed such that the dried and trimmed inflorescences, or buds, of the medical marijuana plant that are collected are representative in maturity and composition of the entire production batch of finished plant material. The sampling timeframe for marijuana buds should be after the completion of the finishing (i.e., drying and trimming) of the plant material production batch.

Homogenization of the finished plant material may be difficult to accomplish prior to sampling due to the heterogeneous nature of the finished plant material. Recommendations from ISO 1839-1980 guidelines for sampling loose leaf tea (i.e., a material similar in nature to cannabis plant material) state that in most cases it is “impracticable and purposeless” to re-blend the contents of a large container of tea in order to obtain a representative sample. USP guidance for sampling articles of botanical origin (USP Chapter <561>) recommends that, for items with component parts larger than 1 cm in any dimension, samples should be withdrawn by hand, then combined and mixed prior to analysis. ISO 1839-1980 also states that if the primary samples consist of loose material, they should be combined to constitute the bulk sample for evaluation.

Quartering is a method to promote the representativeness of a homogenized medical marijuana sample. Quartering involves heaping the adequately mixed and homogenized ground product into a square shape, dividing the heap into four equal quarters, and selecting samples from two of the opposite quarters, which are mixed and sampled (Sexton and Ziskind, 2013; USP Chapter <561>; WHO, 2007). The remaining quarters may then be combined and mixed, then used for microbiological and contaminant testing (Sexton and Ziskind, 2013; USP Chapter <561>; WHO, 2007). The quartering process may be repeated until the required quantity is obtained, and the remaining material may be returned to the batch if possible (USP Chapter <561>; WHO, 2007).

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### ***Solids and semi-solids (Cannabis Resin and Some MIPs)***

Solid and semi-solid products such as resin should be ground and thoroughly mixed, if possible, to be homogenized (USP Chapter <561>; WHO, 2007). A grinding device that minimizes loss (e.g., leaching of resins from finished plant material) should be used, and the grinding device should be cleaned thoroughly after each use. Once ground, quartering, as described above, can be used to collect the sample.

If grinding is impracticable, subsamples of the product should be taken from different areas of the product mass. For example, it might be possible to slice the product mass in sections prior to collection of subsamples or take the subsamples directly from different locations on the product surface (e.g., lower, middle, and upper).

Resin and other solids should not be melted as a means of homogenization. Heating the product may alter the cannabinoid profile or contaminant levels (WHO, 2005) thereby rendering the sample unrepresentative of the source product.

When subsamples are required, subsamples should be composited (combined), if possible, and mixed to obtain a quantity sufficient for evaluation. The quantity sufficient for evaluation may vary by analytical method and laboratory-specific procedures, therefore the RMD should confer with the laboratory to determine the minimum sample quantity required for evaluation.

Compositing subsamples may be impractical for some product types (e.g., hard candies or other products in discrete solid units). In these cases, individual product units can be provided to the laboratory as samples for analysis. In some cases the laboratory may combine extracts or digestates prepared from the solid subsamples and analyze the volumetrically combined extract/digestate as a composite.

### **5.4 Quality Control (QC) Samples**

Duplicate samples shall be collected to provide verification of sampling and laboratory procedures. Specifically, a duplicate should be collected for 5 percent (1 per 20) of the samples collected for each medical marijuana product type. Duplicate samples shall not be identified to the laboratory (this is considered blind quality control). Duplicate samples are used to evaluate any variance in the sampling and analysis procedures.

## **6.0 Sample Collection Procedures**

This section describes sample collection procedures that are generally applicable to any medical marijuana product that RMDs may dispense, including, but not limited to, finished plant material; liquid concentrates or MIPs; resins, waxes, creams, or other semi-solid products; or solid concentrates or MIPs. Because of the wide range of

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medical marijuana products that RMDs may offer, particularly MIPs, these sample collection procedures may require adaptation in some cases. In all cases, sample collection must be conducted in a manner that provides analytically sound and representative samples so that all medical marijuana products dispensed are safe, effective, and accurately labeled. The RMD must document every sampling event and provide this documentation to the Department upon request.

***Prior to Sample Collection.*** The RMD should assemble all equipment and information needed before beginning. Items to assemble before sampling include, but are not limited to, the following:

- Sample collection plan for each product type;
- Logbook or sample collection forms;
- Chain-of-custody forms (COCs);
- Disposable gloves;
- Decontaminated tool(s), such as a spatula, knife, sampling spear, or pipette;
- Stainless steel bowl and implement to homogenize the product (e.g., by stirring, chopping, or grinding);
- Clean, decontaminated surface for sample processing;
- Sample containers appropriate for the analyses required;
- Container labels and pen with indelible ink;
- Supplies to thoroughly clean, decontaminate and dry sampling equipment between samples; and
- A cooler with ice to keep samples cool until refrigeration or shipment to the laboratory.

Sample collection personnel should create a new entry for each sampling event in a sample collection logbook or prepare sample collection forms for documentation of sample collection. Sample collection documentation should identify the sample collection date and start time, participating personnel, a general description of the product type and batch number sampled, a description of the sampling procedures used, and a record of batches that would potentially be impacted should analysis results indicate unacceptable contamination levels.

Sample collection personnel shall identify or determine the cultivation batch number, production batch, and number of samples to be collected based on the guidance provided in Section 5, as well as further guidance obtained in consultation with the laboratory. The number of samples taken from each cultivation and/or production batch must be recorded in the sample collection logbook or forms. Record the sample cultivation and production batch identifiers (ID) for each sample. The batch IDs will be included on sample labels. In addition to the batch ID, create a unique sample ID for

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each sample. Sample identifiers should be unique for a given sample event. Record the batch and sample IDs in the sample collection logbook.

Any tools that contact the samples should be made of stainless steel or other inert material to avoid potential contamination of the sample. Appropriate sample containers should be made of suitable materials.

Preparing sample labels and affixing them to sample containers immediately before sampling is recommended. Information to include on the label includes at a minimum the batch and sample IDs and date/time of collection and by whom. Additional information that must be recorded in documentation, if not on the label, includes sample collector's name, product type, collection method, and other details about the product, such as MIP type or production method.

**Sample Collection.** Collect the planned samples from each cultivation or production batch one at a time. Follow these basic steps for each sample:

1. Wear disposable gloves to mitigate potential for contamination of samples.
2. Ensure that the sampling area is clean and decontaminated and lay out any tools and equipment needed.
3. Collect the sample using an appropriate tool. Do not touch the sample with your hands or allow the sample to touch anything that might cause cross contamination.
4. If necessary, place the sample in the stainless steel bowl or on a decontaminated cutting surface for homogenizing the sample using either the sample collection tool or separate clean, decontaminated implement.
5. Record the time each sample was collected and record any difficulties, inconsistencies with the sampling plan, or other remarks (e.g., environmental conditions) that might be relevant to data analysis or quality assurance.
6. To avoid cross contamination of samples, any tools or equipment that comes in contact with the finished plant material or other marijuana products should be cleaned before collecting the next sample.
7. All samples should be placed in clean, airtight sample containers that are large enough to hold the prescribed sample quantity with minimal headspace. Sample containers must be firmly closed and appropriately labeled.
8. To preserve the chemical and biological composition of the samples, they should be refrigerated or maintained on ice until shipped to the analytical laboratory.
9. Chain-of-custody paperwork should be completed immediately prior to shipment to the analytical laboratory.

Medical marijuana products and MIPs, especially solids or semi-solids such as finished plant material, may be heterogeneous with respect to distribution of cannabinoids or

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contaminants. To obtain a representative sample, liquid products should be thoroughly stirred or mixed before sampling. Solid and semi-solid products must be ground and thoroughly mixed. A grinding device that minimizes loss (e.g., leaching of resins) should be used, and the grinding device should be cleaned thoroughly after each use.

Another method to promote the representativeness of a ground medical marijuana product is quartering. Quartering involves heaping the ground product, dividing the heap into four equal quarters, and selecting samples from two of the quarters, which are combined and mixed (Sexton and Ziskind, 2013). The remaining quarters may then be combined and mixed, then used for microbiological and contaminant testing (Sexton and Ziskind, 2013).

Resin and other solids should not be melted as a means of homogenization. Heating the product may alter the cannabinoid profile or contamination levels (WHO, 2005) thereby rendering the sample unrepresentative of the source product.

Edible products tend to be relatively homogeneous (Sexton and Ziskind, 2013), so a selection of packaged or ready-to-dispense MIPs may be provided to the analytical laboratory to represent a given production batch (Sexton and Ziskind, 2013). MIPs may be either liquid or solid, and the solid MIPs may be of varying density (e.g., baked goods, candies, etc.). Laboratory samples of MIPs shall be homogenized prior to testing such that the sample is representative of the whole product. Homogenized samples should be mixed and quartered similar to the procedure described above. If production batches of individually packaged MIPs are sampled, multiple packaged products should be sampled such that they are representative of the production batch size.

### **7.0 Sample Analysis**

All sample analyses described in this protocol shall be conducted by an independent laboratory that is either:

1. Accredited to International Organization for Standardization (ISO) 17025 by a third party accrediting body such as A2LA or ACLASS, or
2. Certified, registered, or accredited by an organization approved by the Massachusetts Department of Public Health.

Further requirements concerning the eligibility and responsibilities of analytical laboratories are provided in 105 CMR 725.105(C)(2).

In addition to the regulatory qualifications and requirements referenced above, the independent laboratory should have a demonstrated ability to perform the specific

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analytical methods required and to provide defensible documentation and quality assurance.

The sections below identify the analytical methods and analyses required for characterizing the cannabinoid profile of medical marijuana products, as well as the presence and levels of potential contaminants, including metals, pesticides and plant growth regulators, microbiological contaminants and mycotoxins, and residual solvents.

### **7.1 Cannabinoid Profile**

The optimal cannabinoid profile for medical marijuana has not been definitively determined, and this balance may differ depending on the medical condition being treated (AHP, 2013). Although many cannabinoids and related compounds are present in the cannabis plant, characterization of the cannabinoid profile should include, at a minimum, the dry-weight percentage of  $\Delta^9$ -tetrahydrocannabinol ( $\Delta^9$ -THC) and cannabidiol (CBD).

Because target cannabinoid contents and ratios may vary depending on the desired dosage, medical condition, and other use considerations, minimum profile standards are not mandated. However, the cannabinoid profile must be included in product labeling as an aid to patients and caregivers. Analytical procedures for determining cannabinoid profiles are available in AHP (2013).

### **7.2 Metals**

Finished medical marijuana products must be tested for the four metals listed in Exhibit 4. Quantification of metals should be performed with a validated method such as those provided by FDA (2011) or USP (Chapter <233>).

A production batch of finished medical marijuana products (e.g., finished plant material, cannabis resin, or cannabis concentrate) may only be dispensed to patients if all four of the metals are below the upper limits for the respective product in Exhibit 4. These limits are in micrograms ( $\mu\text{g}$ ) of contaminant per kilogram (kg) of product. The upper limits vary by product type because they have been calculated based on equivalent usable marijuana content.

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### Exhibit 4. Analysis Requirements and Recommended Limits for Metals in Finished Medical Marijuana Products

Metal	Upper Limit (µg/kg)		
	Finished Plant Material	Cannabis Resin	Cannabis Concentrate
Arsenic	353	2352	2352
Cadmium	145	964	964
Lead	212	1411	1411
Mercury	701	470	470

### 7.3 Pesticides Residues and Plant Growth Regulators

Non-organic pesticides may not be used to cultivate medical marijuana in Massachusetts (105 CMR 725.105(B)(1)(d)). As discussed in Section 5, all production batches of finished plant material must be tested for residues of prohibited pesticides. At a minimum, samples of finished plant material must be tested for the pesticides, including plant growth regulators, listed in Exhibit 5. These pesticides were identified by AHP (2013) as commonly used in cannabis cultivation. Exhibit 5 identifies appropriate analytical methods for each of the listed pesticides.

Exhibit 5 includes only a small number of the many prohibited non-organic pesticides registered for use in the U.S. To test medical marijuana for pesticides beyond the minimum list in Exhibit 5, Massachusetts recommends additional testing based on the approach USDA Certifying Agents use to analyze prohibited pesticides in organic food. Acknowledging that no method currently exists that analyzes all registered pesticides efficiently (USDA, 2012a), USDA developed a “target” analyte list of 195 prohibited pesticides (USDA, 2011). Under USDA procedures for pesticide residue testing in organic food (USDA, 2013; USDA, 2014), laboratories employed by organic Certifying Agents should “attempt to analyze as many compounds on [the USDA target analyte list] as possible.” Analytical laboratories employed by RMDs for medical marijuana testing in Massachusetts should follow the same approach. Specifically, pesticide testing should be performed consistent with the following sections of *National Organic Program Handbook: Guidance and Instructions for Accredited Certifying Agents and Certified Operations* (USDA, 2014):

- NOP 2611: Laboratory Selection Criteria for Pesticide Residue Testing
- NOP 2611-1: Prohibited Pesticides for NOP Residue Testing
- NOP 2613: Responding to Results from Pesticide Residue Testing

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A further discussion of the application of this testing approach is available in USDA's *2010 - 2011 Pilot Study Pesticide Residue Testing of Organic Produce* (USDA, 2012b)

A production batch of finished plant material may be dispensed to patients or used to make other medical marijuana products if no individual pesticide or plant growth regulator is detected above 10 ppb.

### Exhibit 5. Minimum Analysis Requirements for Residues of Pesticides and Plant Growth Regulators Commonly Used in Cannabis Cultivation

Pesticide	Use	Residue Analytical Methods
Abamectin (Avermectins B1a and B1b)	Insecticide/acaricide	LC-FLD; LC-MS/MS
Acequinocyl	Insecticide/acaricide	LC/MS/MS
Bifenazate	Acaricide	LC; LC-MS/MS
Bifenthrin (synthetic pyrethroid)	Insecticide	GC-ECD; GC-MS/MS
Chlormequat chloride	Plant growth regulator	IC, LC-MS/MS
Cyfluthrin (synthetic pyrethroid)	Insecticide	LC; GC-MS/MS
Daminozide (Alar)	Plant growth regulator	UV spectroscopy; LC-MS/MS
Etoxazole	Acaricide	GC-MS/(MS)
Fenoxycarb	Insecticide	LC/UV; LC-MS/MS
Imazalil	Fungicide	GC-ECD; LC-MS/MS
Imidacloprid	Insecticide	LC-MS/MS
Myclobutanil	Fungicide	GC-ECD; GC-NPD; GSMS/MS; LC-MS/MS
Paclobutrazol	Plant growth regulator; fungicide	LC-MS/MS
Pyrethrins	Insecticide	GC-ECD
Spinosad	Insecticide	LC-MS/MS; Immunoassay
Spiromesifen	Insecticide	GC-MS; LC-MS/MS
Spirotetramat	Insecticide	LC/LC-MS/MS
Trifloxystrobin	Fungicide	GC-NPD; GC-MS/MS; LC-MS/MS

Source AHP (2013)

ECD = Electron capture detector; FLD = Fluorescence detector; GC = Gas chromatography; MS = Mass spectrometry; NPD = Nitrogen phosphorous detector; LC = Liquid chromatography

## 7.4 Microbiological Contaminants and Mycotoxins

Analytical requirements for microbiological contaminants and mycotoxins are listed in Exhibit 6. Requirements for total viable aerobic bacteria, total yeast and mold, total coliforms, and bile-tolerant gram-negative bacteria are given in colony forming unit (CFU) counts per mass of product sample. The requirement for pathogenic *E. coli* and

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Salmonella spp. is based on detection in a 1 gram sample, and the requirement for mycotoxins is based on the concentration per kilogram of sample. Analytical methods for enumerating and identifying specific microbiological contaminants must be consistent with the following United States Pharmacopeia (USP) chapters:

- USP Chapter <61>: Microbiological Examination of Nonsterile Products: Microbial Enumeration Tests. USP 36, Chapter <61>
- USP Chapter <62>: Microbiological Examination of Nonsterile Products: Tests for specified Microorganisms. USP 36, Chapter <62>

Analytical methods for mycotoxins must be consistent with USP chapter:

- USP Chapter <561>: Articles of Botanical Origin. USP 36, Chapter <561>

### Exhibit 6. Analysis Requirements for Microbiological Contaminants and Mycotoxins in Medical Marijuana Products <sup>a</sup>

Cannabis Material	Total Viable Aerobic Bacteria (CFU/g)	Total Yeast and Mold (CFU/g)	Total Coliforms (CFU/g)	Bile-tolerant Gram-negative Bacteria (CFU/g)	E Coli (pathogenic strains) and Salmonella spp.	Mycotoxins <sup>c</sup>
Unprocessed Materials <sup>b</sup>	10 <sup>5</sup>	10 <sup>4</sup>	10 <sup>3</sup>	10 <sup>3</sup>	Not detected in 1 g	<20 µg of any mycotoxin per kg of material
Processed Materials <sup>b</sup>	10 <sup>5</sup>	10 <sup>4</sup>	10 <sup>3</sup>	10 <sup>3</sup>		
CO <sub>2</sub> and Solvent-based Extracts	10 <sup>4</sup>	10 <sup>3</sup>	10 <sup>2</sup>	10 <sup>2</sup>		

CFU: colony forming unit

<sup>a</sup> Except for mycotoxins, analysis requirements are based on AHP (2013).

<sup>b</sup> Unprocessed materials include minimally processed crude cannabis preparations such as inflorescences, accumulated resin glands (kief), and compressed resin glands (hashish). Processed materials include various solid or liquid infused edible preparations, oils, topical preparations, and water-processed resin glands ("bubble hash") (AHP, 2013).

<sup>c</sup> Mycotoxins include aflatoxin B1, aflatoxin B2, aflatoxin O1, aflatoxin O2, and Ochratoxin A.

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### 7.5 Residual Solvents

As discussed in Section 4.2.1, residual solvents testing is required only for cannabis oil. In particular, a production batch of cannabis oil may be dispensed as a finished medical marijuana product or used to make another medical marijuana product only if:

- Laboratory analysis verifies that all solvents used at any stage of cannabis oil production, except in cleaning equipment, are below the limits provided in Exhibit 6; and
- The production batch passes all other applicable testing requirements.

Only solvents listed in Exhibit 7 may be used in the production of cannabis oil. A RMD is required to test only for those solvents used, and it is not required to test for any residual solvents if it can document that no solvents were used in the cannabis oil production process.

The upper limits for residual solvents in Exhibit 7 are given as milligrams of residual solvent per kilogram of cannabis oil. DPH developed the upper limits based on residual solvent standards provided by the United States Pharmacopeia (USP Chapter <467>), the International Conference on Harmonization (ICH, 2011), and AHP (2013). Consistent with the standards provided by these sources, “Class 1” solvents including benzene, carbon tetrachloride, 1,2-dichloroethane, 1,1-dichloroethene, and 1,1,1-trichloroethane may not be used in the production of any medical marijuana product.

Analyses to determine residual solvent concentrations in medical marijuana products must be performed in accordance with the methods identified in USP Chapter <467>.

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**Exhibit 7. Analysis Requirements for Residual Solvents in Cannabis Oil**

<b>Solvent</b>	<b>Upper Limit (mg/kg)</b>
Acetic acid	276.5
Acetone	276.5
Acetonitrile	22.7
Anisole	276.5
1-Butanol	276.5
2-Butanol	276.5
Butyl acetate	276.5
Tert-Butylmethyl ether	276.5
Chlorobenzene	19.9
Chloroform <sup>a</sup>	3.3
Cumene <sup>b</sup>	3.9
Cyclohexane	214.6
1,2-Dichloroethene	103.4
Dichloromethane <sup>a</sup>	33.2
1,2-Dimethoxyethane	5.5
<i>N,N</i> -Dimethylacetamide <sup>a</sup>	60.3
<i>N,N</i> -Dimethylformamide	48.7
Dimethyl sulfoxide	276.5
1,4-Dioxane <sup>a</sup>	21.0
Ethanol	276.5
2-Ethoxyethanol	8.8
Ethyl acetate	276.5
Ethylene glycol	34.3
Ethyl ether	276.5
Ethyl formate	276.5
Formamide	12.2
Formic acid	276.5
Heptane	276.5
Hexane	16.0
Isobutyl acetate	276.5
Isopropyl acetate	276.5
Methanol <sup>a</sup>	165.9
2-Methoxyethanol	2.8
Methyl acetate	276.5
3-Methyl-1-butanol	276.5

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### Exhibit 7. Analysis Requirements for Residual Solvents in Cannabis Oil

Solvent	Upper Limit (mg/kg)
Methylbutylketone	2.8
Methylcyclohexane	65.3
Methylethyl ketone	276.5
Methylisobutyl ketone	276.5
2-Methyl-1-propanol	276.5
N-Methylpyrrolidone <sup>a</sup>	29.3
Nitromethane <sup>a</sup>	2.8
Pentane	276.5
1-Pentanol	276.5
1-Propanol	276.5
2-Propanol	276.5
Propyl acetate	276.5
Pyridine <sup>a</sup>	11.1
Sulfolane	8.8
Tetrahydrofuran	39.8
Tetralin	5.5
Toluene <sup>a</sup>	49.2
1,1,2-Trichloroethylene	4.4
Xylene	120.0

<sup>a</sup> Listed as chemicals known to the State of California to cause cancer or reproductive toxicity under Proposition 65 (CAEPA, 2013, as cited in AHP (2013)).

<sup>b</sup> Cumene is listed as a Class 2 solvent in by ICH (2011) and Class 3 by USP (USP Chapter <467>) and AHP (2013).

## 8.0 Data Evaluation

RMDs are required under 105 CMR 725.105(C)(2)(c) to “have and follow a policy and procedure for responding to results indicating contamination, which shall include destruction of contaminated product and assessment of the source of contamination.”

The analytical results provided by the independent testing laboratory, including those for finished medical marijuana products and MIPs discussed in this protocol, will be a primary means for RMDs to ensure compliance with this requirement.

The independent laboratory results must include, at a minimum, the following in the laboratory data package:

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- Case Narrative:
  - The narrative, written on laboratory letterhead, shall describe any sample receipt, preparation, or analytical issues encountered as well as any method non-conformances or exceedance of QA/QC criteria used by the laboratory.
  - The narrative shall identify the preparation and analytical methods utilized by the laboratory.
  - The narrative shall include a signed statement by an authorized laboratory representative as to the accuracy, completeness, and compliance with the methods of the results presented.
- Chains-of-custody (COC) information or other paperwork indicating requested analyses and documentation of sample collection and receipt.
- Summary of analytical results including sample identifier, methods performed, target analytes analyzed for, result or reporting limit, proper qualifier according to laboratory standard procedures, units of measure, preparation date(s), where applicable, and analysis date(s).

It is highly recommended that the laboratory data package also include sufficient data to evaluate the laboratory results, including a summary of laboratory QA/QC results. The type of applicable QA/QC results differ by analysis method, but can include surrogates or deuterated monitoring compounds, laboratory QC samples such as spikes, blanks, and duplicates, and calibration summaries. It is the responsibility of the RMD to provide information sufficient to demonstrate that the results are accurate and precise, and in line with method capabilities and project data quality objectives (DQOs).

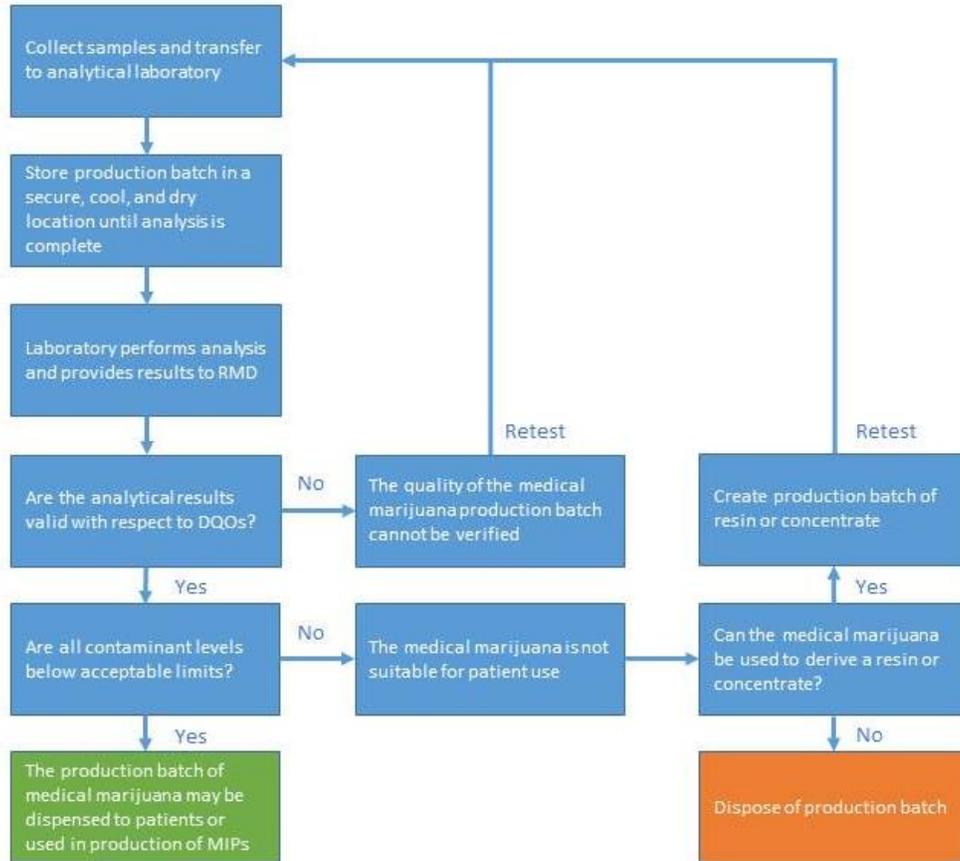
Depending on the outcome of the analysis, the RMD may need to take action to address unacceptable levels of contamination or to perform follow-up investigation. Exhibit 8 is a flowchart the RMD should use to determine the correct course of action in response to each laboratory analytical data package. As discussed above, if any analysis fails to meet all applicable DQOs, then the finished medical marijuana product or MIP cannot be dispensed. In this case, the production batch may be resampled for follow-up testing. A production batch may be retested and records of the original analysis must be retained. If applicable DQOs are not met, the production batch cannot be dispensed to patients or used in the production of MIPs. .

As required by 105 CMR 725.105(C)(2)(b), the RMD must maintain the results of all testing for no less than one year. These records must be available for inspection by the Department, upon request (105 CMR 725.105(I)), and maintained at the RMDs expense in a form and location acceptable to the Department for at least two years after closure (105 CMR 725.105(I)(7)).

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**Exhibit 8. Actions in Response to Laboratory Analytical Results**



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## **Protocol for Sampling and Analysis of Finished Medical Marijuana Products and Marijuana-Infused Products for Massachusetts Registered Medical Marijuana Dispensaries**

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